Synthesis and Biological Evaluation of Novel Paracetamol-Triazole Conjugates

Necla KULABAŞ^{*}, Merve GÜRBOĞA^{**}, Özlem BİNGÖL ÖZAKPINAR^{***}, Jianyang LIU^{****}, Per-Johan JAKOBSSON^{*****}, Özkan DANIŞ^{******}, Ayşe OGAN^{*******}, İlkay KÜÇÜKGÜZEL^{********}

Synthesis and Biological Evaluation of Novel Paracetamol-Triazole Conjugates

SUMMARY

Some new triazole containing acetamide derivatives **9-20** using paracetamol as starting material were synthesized, and their structures were verified by FTIR, NMR (¹H and ¹³C) and mass spectral data. Compounds **9-20** were tested against five human cancer cell lines (lung cancer A549, chronic myelogenous leukemia K562, breast cancer MCF-7, prostate cancer PC-3, neuroblastoma SH-SY5Y cell lines) for in vitro cytotoxic activities. They were also evaluated their cytotoxic effect on mouse embryonic fibroblast cells (NIH/3T3) to define selectivity by MTT assay. Additionally, we evaluated the potential mPGES-1 and COX-1/2 inhibitory effect of twelve target compounds **9-20**. While none of the synthesized compounds exhibited significant inhibition against both cancer cells and mPGES-1 as well as COX-1/2, it was determined that they were not cytotoxic against healthy cells, too. Finally, ADMET properties of newly synthesized compounds were estimated using in silico methods.

Key Words: Paracetamol, 1,2,4-triazole, cancer, mPGES-1, COX-1/2. Yeni Parasetamol-Triazol Konjugatlarının Sentezi ve Biyolojik Değerlendirmesi

ÖΖ

Başlangıç maddesi olarak parasetamol kullanılarak bazı yeni triazol içeren asetamid türevleri **9-20** sentezlendi ve yapıları FTIR, NMR (¹H and ¹³C) ve kütle spektral verileri ile karakterize edildi. Beş insan kanser hücre hattına (akciğer kanseri A549, kronik miyelojenöz lösemi K562, meme kanseri MCF-7, prostat kanseri PC-3, nöroblastoma SH-SY5Y hücre hatları) karşı sentezlenen tüm moleküllerin in vitro sitotoksik aktiviteleri incelendi ve ayrıca seçiciliği tanımlamak için fare embriyonik fibroblast hücreleri (NIH/3T3) üzerinde sitotoksik etkileri MTT yöntemiyle test edildi. Ek olarak, on iki hedef bileşik **9-20**, mPGES-1 ve COX-1/2 inhibe edici etkileri açısından tarandı. Sentezlenen bileşiklerin hiçbiri hem kanser hücrelerine hem de mPGES-1 ve COX-1/2 enzimlerine karşı anlamlı bir inhibisyon göstermezken, sağlıklı hücrelere karşı da sitotoksik olmadıkları belirlendi. Son olarak yeni sentezlenen bileşiklerin ADMET özellikleri in siliko yöntemler kullanılarak tahmin edildi.

Anahtar Kelimeler: Parasetamol, 1,2,4-triazol, kanser, mPGES-1, COX-1/2.

Received: 7.08.2023 Revised: 23.08.2023 Accepted: 24.08.2023

* ORCID: 0000-0003-2273-5094, Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Marmara University, Başıbüyük, İstanbul, Türkiye

" ORCID: 0000-0003-4614-7094, Department of Biochemistry, Faculty of Pharmacy, Marmara University, Başıbüyük, İstanbul, Türkiye

" ORCID: 0000-0003-0287-563, Department of Biochemistry, Faculty of Pharmacy, Marmara University, Başıbüyük, İstanbul, Türkiye

"" ORCID: 0000-0002-8683-1109, Rheumatology Division, Department of Medicine, Solna, Karolinska Institutet & Karolinska University Hospital, Stockholm, Sweden "" ORCID: 0000-0001-7665-9715, Rheumatology Division, Department of Medicine, Solna, Karolinska Institutet & Karolinska University Hospital, Stockholm, Sweden "" ORCID: 0000-0003-1781-0520, Department of Chemistry, Faculty of Science, Marmara University, İstanbul, Türkiye

- ORCID: 0000-0002-8973-9762, Department of Chemistry, Faculty of Science, Marmara University, İstanbul, Türkiye
- """ ORCID: 0000-0002-7188-1859 Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Fenerbahçe University, Ataşehir, İstanbul, Türkiye

° Corresponding Author; İlkay KÜÇÜKGÜZEL

Tel: +90 216 910 1907 , E-mail: kucukguzel@hotmail.com, ilkay.kucukguzel@fbu.edu.tr

INTRODUCTION

Characterized by uncontrolled cell division, cancer is one of the significant causes of death worldwide. According to the World Health Organization (WHO) data, cancer caused about 10 million deaths (nearly one in six deaths) in 2020 (WHO, 2022). Today, the primary method of treating cancer is surgery-based therapy, but patients undergoing surgical treatment are often at risk of death and perioperative complications. Apart from surgical interventions, radiotherapy and chemotherapy, which are known to inhibit tumor cell growth, proliferation and metastasis, play a critical role in cancer treatment. However, the used drugs can cause more problems due to their undesired side effects (Zhai et al., 2022). Hence, ongoing research endeavors are focused on identifying novel anticancer agents characterized by improved side effect profiles.

Cancer formation, development, and metastasis are closely linked to the tumor microenvironment, and clinical studies have reported increasing levels of microsomal prostaglandin E2 synthase-1 (mPGES-1) and cyclooxygenase-2 (COX-2) in several human cancers (Murakami, 2011) bind to their cognate G protein-coupled receptors (GPCRs). Even the relationship of breast (Howe et al., 2013), colon (Sasaki, Nakatani, & Hara, 2015), prostate (Hanaka et al., 2009), lung (Chang et al., 2012), neuroblastoma (Kock et al., 2018) and many more cancer types with COX-2 and mPGES-1 enzymes has been reported in the literature. Cyclooxygenases (COXs) are ratelimiting enzymes, which are involved in the conversion of arachidonic acid to prostaglandin E2 (PGE2). It has two structurally analogous isoforms (COX-1 and COX-2). Long-term use of COX-1 and COX-2 inhibitors has been reported to beget gastrointestinal and cardiovascular side effects, respectively (Crofford, 1997). It is known that COX-1 has also been known to be regulated, for example, Interleukin 4 (IL-4) and results in increased production of PGs (Shay et al., 2017), and the excessive of PGE2 is synthesized by COX-2 enzyme and mPGES-1 enzyme (Akasaka, So, & Ruan, 2015). Therefore, mPGES-1 and COX-396

1/2 enzymes have been the focus of attention as new targets in cancer therapy. The triazole ring is an electron-rich aromatic structure. It exhibits a wide range of biological activities such as anti-tuberculosis (Kücükgüzel, Kücükgüzel, Rollas, & Kiraz, 2001), anti-inflammatory (Küçükgüzel et al., 2007) and anticancer activities (Kulabaş et al., 2016) due to their ability to easily bind to different macromolecular targets with weak interactions. Moreover, many 1,2,4-triazole-thioether derivatives were evaluated as anticancer (Demirbolat et al., 2022; Kulabaş et al., 2016; Zengin, Unsal Tan, Arafa, & Balkan, 2022) agents whereas several reports indicate them as potential inhibitors of mPGES-1 (Bülbül et al., 2022; Erensoy et al., 2023; He, Li, Liu, & Lai, 2013). Paracetamol, a non-opioid antipyretic and analgesic agent, is used in the treatment of fever and pain as it inhibits COX-1/2 enzymes (Graham, Davies, Day, Mohamudally, & Scott, 2013). In light of previous studies on the anticancer and anti-inflammatory effect of triazolyl-sulfanyl acetamides, as shown in Figure 1, we decided to design novel paracetamoltriazole conjugates bearing acetanilide moiety.

MATERIAL AND METHOD

Chemicals and instruments

All solvents and reagents utilized in this study were of analytical grade and were procured from commercially reputable sources. The compounds' purity was ascertained through thin-layer chromatography (TLC) conducted on Merck silica gel 60 F254 aluminum sheets (Merck, Darmstadt, Germany). UV light at a wavelength of λ = 254 nm was employed to visualize the spots. The determination of all melting points (°C, uncorrected) was carried out utilizing the Electro thermal IA9300 melting point apparatus. Infrared spectra (FTIR) were captured using a Shimadzu FTIR 8400s, with data presented in wavenumbers (cm⁻¹). Nuclear Magnetic Resonance (NMR) spectra were acquired employing a Bruker Avance NMR spectrometer operating at frequencies of 300 MHz for ¹H NMR and 75 MHz for ¹³C NMR.



Figure 1. The reported 1,2,4-triazoles with antiproliferative and mPGES-1 inhibitory effects and our designed paracetamol-triazole conjugates 9-20.

Chemical shifts were denoted in parts per million (ppm), referenced downfield from tetramethylsilane (TMS), with DMSO-d₆ serving as the solvent. The mass spectra were obtained using the Shimadzu LC-MS/MS-8030 system (Shimadzu Corporation, Kyoto, Japan). Operating within the realm of positive electrospray ionization (ESI) mode, the instrument entailed a nebulizing gas flow rate of 2.8 L/min, a drying gas flow of 15 L/min, a DL temperature of 250 °C, a heat block temperature of 400 °C, CID gas at 230 kPa, collision energy set at -5.0 V, a scan speed of 15000 u/sec, and an event time of 0.030 sec. Elemental analyses were conducted using the Leco CHNS 932 apparatus.

Chemistry

method General synthesis of N-aryl-2chloroacetamides (I-IV)

The reaction mixtures consisted of dissolved substituted aniline derivatives (0.003 mol) in dichloromethane (DCM), followed by the addition of trimethylamine (TEA) (0.5 ml). 2-Chloroacetyl chloride (0.005 mol) was dropped onto the mixture at 0-5°C. Then the flask content was stirred under reflux at 50°C for 4 h. After completion of the reaction according to TLC; the mixtures were cooled, filtered, dried, and recrystallized from methanol to obtain products I-IV (Demirci et al., 2018; Kulabaş et al., 2022).

2-Chloro-N-(4-acetylphenyl)acetamide (I): M.p. 146.2-147.4°C (lit. 169.8-172 °C, (Haider & Hamada, 2012)), yield 85.9%. TLC Rf: 0.36. IR cm⁻¹: 3281 (N-H stretching band (str)), 1703 (C=O str, ketone), 1651 (C=O str, anilide).

2-Chloro-N-(4-nitrophenyl)acetamide (II): M.p. 176.2-177 °C (lit. 177-180 °C, (Zhaowen et al., 2007)), yield 70%. TLC Rf: 0.66. IR cm⁻¹: 3266 (N-H str), 1684 (C=O str, anilide).

2-Chloro-N-(2-methyl-5-nitrophenyl)acetamide (III): M.p. 155.5-155.7 °C (lit. 144-146 °C, (Svetkin, Minlibaeva, & Khafizova, 1961)), yield 63%. TLC Rf: 0.59. IR cm⁻¹: 3252 (N-H str), 1672 (C=O str, anilide).

Ethyl 4-[(chloroacetyl)amino]benzoate (IV): M.p. 112.4-112.7 °C (MeOH) (lit. 106-107 °C,(Haider & Hamada, 2012)), yield 78%. TLC Rf: 0.67. IR cm⁻¹: 3273 (N-H str), 1717 (C=O str, ester), 1674 (C=O str, anilide).

General synthesis method of ethyl (4-acetamidophenoxy)acetate (1)

Paracetamol (0.05 mol) and 0.075 mol anhydrous potassium carbonate were dissolved in dry acetone and stirred at 65 °C for 4 h under reflux. Then ethyl 2-bromoacetate (0.055 mol) was added dropwise to this mixture during 1 h, and refluxed for another 8 h. Following the termination of the reaction, the mixture underwent filtration, and the excess acetone was subjected to distillation. Afterward, the resulting product underwent a washing process with water, followed by drying and recrystallization from ethanol (Küçükgüzel, Tatar, Küçükgüzel, Rollas, & De Clercq, 2008).

Ethyl [4-(acetylamino)phenoxy]acetate (1): M.p. 103.2-103.6 °C (MeOH) (lit. 106-107 °C,(Morar, Cost, Lameiras, Antheaume, & Darabantu, 2015)), yield 68.5%. TLC Rf: 0.50. IR cm⁻¹: 3379 (N-H), 1738 (C=O, ester), 1674 (C=O, anilide).

General synthesis method of 2-(4-acetamidophenoxy) acetohydrazide (2)

The ethanol solution containing compound **1** (0.015 mol) was subjected to reflux with hydrazine monohydrate (0.045 mol) for 3 h. After cooling the reaction mixture, the resulting residue was washed with ice-cold water and subsequently filtered. Compound **2** was acquired by recrystallization from ethanol (Küçükgüzel et al., 2008).

N-[4-(2-Hydrazinyl-2-oxoethoxy)phenyl] acetamide (**2**): M.p. 193.5-194.1°C (MeOH) (lit. 194-195 °C,(Nargund, Reddy, & Hariprasad, 1994)), yield 85.6%. TLC Rf: 0.30. IR cm⁻¹: 3322, 3298, 3190 (N-H), 1676 (C=O, anilide), 1642 (C=O, hydrazide).

General synthesis method of N-methyl/ ethyl/phenyl-2-(substituted aryloxyacetyl) hydrazinecarbothioamide (3-5)

Compound **2** (0.03 mol) was mixed with methyl/ ethyl/phenylisothiocyanate (0.03 mol) in methanol (40 ml), and this mixture was refluxed for 6 h at 95°C. **398** The cooled solution was filtered and then washed with water. The resulting solid product was subjected to recrystallization from methanol, resulting in the acquisition of compounds **3-5** (Kulabaş et al., 2016).

N-(4-{2-[2-(Methylcarbamothioyl)hydrazinyl]-2oxoethoxy}phenyl)acetamide (**3**): M.p. 195.4-196.8 °C (MeOH) (lit. 222-226 °C,(İlkay Küçükgüzel et al., 2008)), yield 62%. TLC Rf: 0.48. IR *v* (cm⁻¹): 3223, 3131, 3072 (N-H str), 2953-2870 (C-H str), 1720, 1640 (C=O str), 1229 (C=S str).

N-(4-{2-[2-(Ethylcarbamothioyl)hydrazinyl]-2oxoethoxy}phenyl)acetamide (**4**): M.p. 191.5-192.1°C (MeOH) (lit. 182-184 °C,(İlkay Küçükgüzel et al., 2008)), yield 60%. TLC Rf: 0.57. IR *v* (cm⁻¹): 3264, 3131 (N-H str), 3070-2918 (C-H str), 1690, 1643 (C=O str), 1215 (C=S str).

N-(4-{2-[2-(Phenylcarbamothioyl)hydrazinyl]-2oxoethoxy}phenyl)acetamide (5): M.p. 196.7-196.9 °C (MeOH) (lit. 190-192 ° C,(İlkay Küçükgüzel et al., 2008)), yield 51%. TLC Rf: 0.07. IR v (cm⁻¹): 3295, 3243, 3129 (N-H str), 3101-2906 (C-H str), 1699, 1661 (C=O str), 1238 (C=S str).

General synthesis method of 4-methyl/ethyl/ phenyl-5-(substituted aryloxymethyl)-2,4-dihydro-3H-1,2,4-triazole-3-thione (6-8)

A mixture of compounds **3**, **4**, or **5** (0.02 mol) and TEA (8 ml) in ethanol (40 ml) was refluxed for 6 h at 95 °C. After the reaction was terminated, the flask content was neutralized by 5% HCl to give desired compounds. The obtained solid product was subjected to filtration, followed by drying and recrystallization from ethanol (Kulabaş et al., 2016).

N-{4-[(4-Methyl-5-thioxo-4,5-dihydro-1*H*-1,2,4-triazol-3-yl)methoxy]phenyl}acetamide (**6**): M.p. 221.7-211.9 °C, yield 71%. TLC Rf: 0.26. IR *ν* (cm⁻¹): 3289, 3133 (N-H str), 3055-2949 (C-H str), 1669 (C=O str), 1539 (C=N str), 1275 (C=S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.06 (3H, t, *J*= 6 *Hz*, C**H**₃CH₂OH); 2.01 (3H, s, -COC**H**₃); 2.33 (3H, s, Ar-C**H**₃); 3.49 (3H, s, N-C**H**₃); 5.17 (2H, s, -OC**H**₂); 7.01 (2H, d, J= 9 Hz, Ar-**H**); 7.51 (2H, d, *J*= 9 Hz, Ar-

H); 9.86 (1H, s, amide -N**H**-); 13.83 (1H, s, triazole -N**H**-). Anal. calc. for $C_{12}H_{14}N_4O_2S.1/3$ mol C_2H_5OH C: 51.80, H: 5.49, N: 19.08, S: 10.92; found C: 51.78, H: 5.07, N: 20.13, S: 11,52.

N-{4-[(4-Ethyl-5-thioxo-4,5-dihydro-1*H*-1,2,4triazol-3-yl)methoxy]phenyl}acetamide (7): M.p. 198.4-198.8 °C, yield 69%. TLC Rf: 0.33. IR ν (cm⁻¹): 3349 (N-H str), 3084-2851 (C-H str), 1672 (C=O str), 1539 (C=N str), 1275 (C=S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.06 (3H, t, *J*= *7 Hz*, CH₃CH₂OH); 1.31 (3H, t, *J*= *7.2 Hz*, N-CH₂CH₃); 2.01 (3H, s, -COCH₃); 3.39-3.49 (m, CH₃CH₂OH and DMSO); 4.04 (2H, q, *J*= 6.6 *Hz* and *7 Hz*, N-CH₂CH₃); 4.39 (1H, t, *J*= 5.1 Hz, CH₃CH₂OH); 5.18 (2H, s, -OCH₂); 7.00 (2H, d, *J*= 9 *Hz*, Ar-H); 7.51 (2H, d, *J*= 9 *Hz*, Ar-H); 9.86 (1H, s, amide -NH-); 13.84 (1H, s, triazole -NH-). Anal. calc. for C₁₃H₁₆N₄O₂S.1/2 mol C₂H₅OH C: 53.31, H: 6.07, N: 17.76, S: 10.17; found C: 53.48, H: 5.52, N: 19.16, S: 10.97.

N-{4-[(4-Phenyl-5-thioxo-4,5-dihydro-1*H*-1,2,4triazol-3-yl)methoxy]phenyl}acetamide (**8**): M.p. 213.9-214.1 °C, yield 63%. TLC Rf: 0.38. IR v (cm⁻¹): 3293 (N-H str), 3094-2920 (C-H str), 1659 (C=O str), 1543 (C=N str), 1240 (C=S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.99 (3H, s, -COCH₃); 4.91 (2H, s, -OCH₂); 6.75 (2H, d, J= 9 Hz, Ar-H); 7.38-7.54 (7H, m, Ar-H); 9.81 (1H, s, amide -NH-); 14.06 (1H, s, triazole -NH-). Anal. calcd. for $C_{17}H_{16}N_4O_2S$ C: 59.62, H: 4.48, N: 16.30, S: 9.29; found C: 59.98, H: 4.74, N: 16.46, S: 9.42.

General synthesis method of 2-{[3-(substituted aryloxymethyl)-4-methyl/ethyl/phenyl-4,5dihydro-1*H*-1,2,4-triazol-5-yl]sulfanyl}-*N*-(4substituted phenyl)acetamide (9-20)

The solution of compounds **6**, 7, or **8** (0.001 mol) was prepared in dimethylformamide (DMF) (10 ml), and subsequently, anhydrous potassium carbonate (0.002 mol) was introduced into the solution. Following an hour of stirring at room temperature, the solution containing compounds **I-IV** (0.001 mol) in DMF (3 ml) was introduced into the mixture.

The reaction medium was stirred by refluxing for 6 h at 50-60 °C. After the reaction was terminated, the solution was cooled, and the product was filtered, and subjected to washing with water. Finally, compounds **9-20** were recrystallized from ethanol (Kulabaş et al., 2016).

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4-methyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(4acetylphenyl)acetamide (9): M.p. 280.9-281.8 °C, yield 50%. TLC Rf: 0.25. IR v (cm⁻¹): 3237, 3187 (N-H str), 3102-2863 (C-H str), 1694, 1659 (C=O str), 1541 (C=N str), 671 (C-S str). ¹H NMR (300 MHz, DMSO_d6) δ ppm: 2.01 (3H, s, -COCH₂); 2.53 (3H, s, -COCH₃); 3.61 (3H, s, N-CH₃); 2.73 and 2.89 (s, DMF peak); 4.16 (2H, s, -S-CH, -); 5.23 (2H, s, -OCH,); 7.00 (2H, d, *J*= 9 *Hz*, Ar-**H**); 7.49 (2H, d, *J*= 9 *Hz*, Ar-**H**); 7.70 (2H, d, J= 9 Hz, Ar-H); 7.95 (2H, d, J= 9 Hz, Ar-H); 9.83 (1H, s, -NH-); 10.67 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d_c/TMS): δ ppm: 24.28, 26.90, 30.93 (aliphatic C), 37.94 (S-CH₂), 60.79 (O-CH₂), 115.59, 118.87, 120.88, 130.02, 132.43, 133.95, 143.50, 151.01 (Ar-C), 152.67 and 153.71 (triazole C3 and C5), 166.77 and 168.70 (CO, amide), 196.97 (Ar-COCH₂). Anal. calc. for C₂₂H₂₂N₅O₄S C:58.26, H:5.11, N:15.44, S:7.07; found C:58.06, H:5.25, N:15.37, S:7.07. LR-MS (ESI) (m/z): calc. for (M+Na)⁺: 476.15, found: 476.15. Calc. for (M-H): 452.15, found: 452.15.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4-methyl-4*H*-1,2,4-triazol-3-yl)sulfanyl]-*N*-(4nitrophenyl)acetamide (**10**): M.p. 279.5-280.0 °C, yield 58%. TLC Rf: 0.29. IR *v* (cm⁻¹): 3219, 3159 (N-H str), 3096-2834 (C-H str), 1694, 1659 (C=O str), 1547 (C=N str), 688 (C-S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 2.01 (3H, s, -COCH₃); 2.73 and 2.89 (s, DMF peak); 3.62 (3H, s, N-CH₃); 4.19 (2H, s, -S-CH₂-); 5.23 (2H, s, -OCH₂); 7.00 (2H, d, *J*= 9 *Hz*, Ar-H); 7.49 (2H, d, *J*= 9 *Hz*, Ar-H); 7.81 (2H, d, *J*= 9 *Hz*, Ar-H); 8.23 (2H, d, *J*= 9 *Hz*, Ar-H); 9.83 (1H, s, -NH-); 10.94 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/ TMS): δ ppm: 19.03, 24.25 (aliphatic **C**), 37.32 (S-CH₂), 60.58 (O-CH₂), 115.47, 119.32, 120.79, 125.54, 127.43, 130.28, 130.62, 133.03, 133.88, 142.86, 145.34, 151.83 (Ar-C), 152.33 and 153.47 (triazole C3 and C5), 167.00 and 168.27 (CO, amide). Anal. calc. for $C_{20}H_{20}N_6O_5S.1/5$ mol DMF C:52.52, H:4.58, N:18.43, S:6.81; found C:51.24, H:4.55, N:18.12, S:6.94. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 479.12, found: 479.10. Calc. for (M-H)⁻: 455.12, found: 455.10.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4methyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(2-methyl-5-nitrophenyl)acetamide (11): M.p. 216.4-217.7 °C, yield 47%. TLC Rf: 0.27. IR v (cm⁻¹): 3243, 3190 (N-H str), 3026-2834 (C-H str), 1669 (C=O str), 1537 (C=N str), 681 (C-S str). ¹H NMR (300 MHz, DMSO-d_z) δ ppm: 1.06 (0.3 H, t, *J*= 6 *Hz*, 0.1 mol CH₂CH₂OH); 2.01 (3H, s, -COCH₃); 2.33 (3H, s, Ar-CH₃); 3.62 (3H, s, N-CH₂); 4.21 (2H, s, -S-CH₂-); 5.24 (2H, s, -OCH₂); 7.01 (2H, d, J= 9 Hz, Ar-H); 7.46-7.52 (3H, m, Ar-H); 7.95 (1H, dd, *J*= 2.4 Hz and 9 Hz, Ar-H); 8.50 (1H, d, *J*= 2.4 Hz, Ar-H); 9.83 (1H, s, -NH-); 9.99 (1H, s, -NH-). ¹³C NMR (75 MHz, DMSO-d_c/TMS): δ ppm: 18.52, 24.28, 30.96 (aliphatic C), 37.23 (S-CH₂), 60.76 (O-CH₂), 115.57, 118.25, 119.90, 120.88, 131.92, 133.95, 137.41, 139.07, 146.18, 151.20 (Ar-C), 152.72 and 153.70 (triazole C3 and C5), 167.21 and 168.30 (CO, amide). Anal. calc. for $C_{21}H_{22}N_6O_5S.0.1$ mol C₂H₂OH C: 53.59, H:4.79, N:17.69, S:6.75; found C:53.66 H:4.75, N:17.38, S:6.87. LR-MS (ESI) (m/z): Calc. for (M+Na)+: 493.14, found: 493.15. Calc. for (M-H)-: 469.14, found: 469.15.

Ethyl 4-[({[5-{[4-(acetylamino)phenoxy]methyl}-4-methyl-4*H*-1,2,4-triazol-3-yl]sulfanyl}acetyl) amino]benzoate (**12**): M.p. 241.0-241.3 °C, yield 56%. TLC Rf: 0.33. IR *ν* (cm⁻¹): 3256, 3244 (N-H str), 3107-2932 (C-H str), 1686 (C=O str), 1525 (C=N str), 635 (C-S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.31 (3H, t, *J* = 6 *Hz*, CH₃CH₂O-); 2.01 (3H, s, -COCH₃); 3.61(3H, s, N-CH₃); 4.16 (2H, s, -S-CH₂-); 4.29 (2H, q, *J* = 7.2 *Hz*, CH₃CH₂O-); 5.22 (2H, s, -OCH₂); 7.00 (2H, d, J= 9 Hz, Ar-H); 7.49 (2H, d, *J* = 9 *Hz*, Ar-H); 7.70 (2H, d, *J* = 9 *Hz*, Ar-H); 7.93 (2H, d, *J* = 9 *Hz*, Ar-H); 9.83 (1H, s, -NH-); 10.67 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/TMS): δ ppm: 14.67, 24.28, 30.92 (aliphatic C), 37.91 (S-CH₂), 60.94 (O-CH₂), 115.58, **400** 118.98, 120.88, 125.00, 130.79, 133.95, 143.52, 151.02 (Ar-C), 152.66 and 153.71 (triazole C3 and C5), 165.73 (CO, ester), 166.78 and 168.29 (CO, amide). Anal. calc. for $C_{23}H_{25}N_5O_5S$ C: 57.13, H:5.21, N:14.48, S:6.63; found %C:56.54, H:4.82, N:14.40, S:6.58. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 506.16, found: 506.15. Calc. for (M-H)⁻: 482.16, found: 482.20.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4-ethyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(4acetylphenyl)acetamide (13): M.p. 262.1-262.8 °C, yield 61%. TLC Rf: 0.39. IR v (cm⁻¹): 3241, 3188 (N-H str), 3119-2843 (C-H str), 1661 (C=O str), 1541 (C=N str), 662 (C-S str). ¹H NMR (300 MHz, DMSO-d_z) δ ppm: 1.31 (3H, t, J= 6 Hz, N-CH₂CH₃); 2.01 (3H, s, -COCH₂); 2.73 and 2.89 (s, DMF peak); 4.06 (2H, q, J= 6 Hz and 9 Hz, N-CH, CH,); 4.24 (2H, s, -S-CH, -); 5.24 (2H, s, -OCH₂); 7.00 (2H, d, *J*= 9 *Hz*, Ar-H); 7.50 (2H, d, J= 9 Hz, Ar-H); 7.70 (2H, d, J= 9 Hz, Ar-**H**); 7.95 (2H, d, *J*= 9 *Hz*, Ar-**H**); 9.84 (1H, s, -N**H**-); 10.70 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₂/ TMS): δ ppm: 15.53, 24.27, 26.90 (aliphatic C), 37.86 (S-CH₂), 60.51 (O-CH₂), 115.37, 118.88, 120.96, 130.01, 132.40, 133.89, 143.57, 150.54 (Ar-C), 152.09 and 153.62 (triazole C3 and C5), 166.70 and 168.30 (CO, amide), 196.97 (Ar-COCH₂). Anal. calc. for C₂₃H₂₅N₅O₄S C:59.08, H:5.39, N:14.98, S:6.86; found C:58.46, H:5.22, N:14.79, S:6.71. LR-MS (ESI) (m/z): Calc. for (M+Na)+: 490.16, found: 490.20. Calc. for (M-H): 466.16, found: 466.10.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4-ethyl-4*H*-1,2,4-triazol-3-yl)sulfanyl]-*N*-(4nitrophenyl)acetamide (14): M.p. 272.9-273.9 °C, yield 62%. TLC Rf: 0.30. IR v (cm⁻¹): 3217 (N-H str), 3055-2857 (C-H str), 1697, 1653 (C=O str), 1574 (C=N str), 665 (C-S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.31 (3H, t, *J*= 6 *Hz*, N-CH₂CH₃); 2.01 (3H, s, -COCH₃); 2.73 and 2.89 (s, DMF peak); 4.06 (2H, q, *J*= 6 *Hz* and 9 *Hz*, N-CH₂CH₃); 4.26 (2H, s, -S-CH₂-); 5.24 (2H, s, -OCH₂); 7.00 (2H, d, *J*= 9 *Hz*, Ar-H); 7.49 (2H, d, *J*= 9 *Hz*, Ar-H); 7.82 (2H, d, *J*= 9 *Hz*, Ar-H); 8.24 (2H, d, *J*= 9 *Hz*, Ar-H); 9.83 (1H, s, -NH-); 10.97 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/TMS): δ ppm: 24.27, 30.93 (aliphatic C), 37.90 (S-CH₂), 60.78 (O-CH₂), 115.58, 119.32, 120.87, 125.53, 133.95, 142.88, 145.31, 150.94 (Ar-C), 152.69 and 153.69 (triazole C3 and C5), 167.22 and 168.29 (CO, amide). Anal. calc. for $C_{21}H_{22}N_6O_5S$ C: 53.61, H:4.71, N:17.86, S:6.82; found C:53.53, H:4.50, N:17.44, S:6.74. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 493.14, found: 493.15. Calc. for (M-H)⁻: 469.14, found: 469.15.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4ethyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(2-methyl-5-nitrophenyl)acetamide (15): M.p. 207.6-208.5 °C, yield 45%. TLC Rf: 0.36. IR v (cm⁻¹): 3237, 3187 (N-H str), 3134-2843 (C-H str), 1667 (C=O str), 1534 (C=N str), 664 (C-S str). ¹H NMR (300 MHz, DMSO- d_{6}) δ ppm: 1.06 (0.3H, t, J= 6 Hz, 0.1 mol CH, CH, OH); 1.31 (t, 3H, *J*= 6 *Hz*, N-CH₂CH₃); 2.01 (3H, s, -COCH₃); 2.34 (3H, s, Ar-CH₃); 4.06 (2H, q, J= 6 Hz and 9 Hz, N-CH₂CH₃); 4.27 (2H, s, -S-CH₂-); 5.25 (2H, s, -OCH₂); 7.01 (2H, d, J= 9 Hz, Ar-H); 7.48-7.52 (3H, m, Ar-H); 7.95 (1H, dd, J= 2.7 Hz and 8.1 Hz, Ar-**H**); 8.50 (1H, d, *J*= 2.7 *Hz*, Ar-**H**); 9.84 (1H, s, -N**H**-); 10.02 (1H, s, -NH-). ¹³C NMR (75 MHz, DMSO-d_c/ TMS): δ ppm: 15.52, 18.51, 19.03, 24.27 (aliphatic C), 37.17 (S-CH₂), 60.47(O-CH₂), 115.37, 118.29, 119.91, 120.96, 131.92, 133.89, 137.42, 139.09, 146.18, 150.68 (Ar-C), 152.18 and 153.61 (triazole C3 and C5), 167.16 and 168.30 (CO, amide). Anal. calc. for C₂₂H₂₄N₆O₅S C: 54.53, H:4.99, N:17.34, S:6.62; found C:54.43 H:5.17, N:17.13, S:6.66. LR-MS (ESI) (m/z): Calc. for (M+Na)+: 507.15, found: 507.20. Calc. for (M-H): 483.15, found: 483.20.

Ethyl 4-[({[5-{[4-(acetylamino)phenoxy]methyl}-4-ethyl-4*H*-1,2,4-triazol-3-yl]sulfanyl}acetyl) amino] benzoate (**16**): M.p. 242.1-243.3 °C, yield 55%. TLC Rf: 0.41. IR *v* (cm⁻¹): 3258, 3181 (N-H str), 3151-2955 (C-H str), 1696, 1659 (C=O str), 15413 (C=N str), 662 (C-S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.31 (6H, t, *J* = 6 *Hz*, CH₃CH₂O- and N-CH₂CH₃); 2.01 (3H, s, -COCH₃); 2.73 and 2.89 (s, DMF peak); 4.05 (2H, q, *J* = 6 *Hz* and *9 Hz*, N-CH₂CH₃); 4.23-4.32 (4H, m, -S-CH₂- and CH₃CH₂O-); 5.24 (2H, s, -OCH₂); 7.00 (2H, d, *J* = 9 *Hz*, Ar-H); 7.50 (2H, d, *J* = 9 *Hz*, Ar-H); 7.71 (2H, d, J = 9 Hz, Ar-H); 7.93 (2H, d, J = 9 Hz, Ar-H); 9.84 (1H, s, -NH-); 10.70 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/TMS): δ ppm: 14.67, 15.53, 24.27 (aliphatic C), 37.81 (S-CH₂), 60.93 (O-CH₂), 115.37, 118.98, 120.96, 124.99, 130.78, 133.89, 143.53, 150.54 (Ar-C), 152.09 and 153.63 (triazole C3 and C5), 165.73 (CO, ester), 166.68 and 168.29 (CO, amide). Anal. calc. for $C_{24}H_{27}N_5O_5S$ C: 59.08, H:5.39, N:14.98, S:6.86; found C:58.46, H:5.22, N:14.79, S:6.71. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 520.17, found: 520.20. Calc. for (M-H)⁻: 496.17, found: 496.20.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4phenyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(4-acetyl phenyl)acetamide (17): M.p. 238.1-239.0 °C, yield 52%. TLC Rf: 0.25. IR v (cm⁻¹): 3256, 3183 (N-H str), 3100-2930 (C-H str), 1672, 1667 (C=O str), 1541 (C=N str), 667 (C-S str). ¹H NMR (300 MHz, DMSO- d_6) δ ppm: 1.99 (3H, s, -COCH₃); 2.53 (3H, s, -COCH₃); 4.24 (2H, s, -S-CH₂-); 5.03 (2H, s, -OCH₂); 6.79 (2H, d, J= 9 Hz, Ar-H); 7.41 (2H, d, J= 9 Hz, Ar-H); 7.50-7.58 (5H, m, Ar-H); 7.71 (2H, d, J= 9 Hz, Ar-H); 7.95 (2H, d, *J*= 9 *Hz*, Ar-H); 9.79 (1H, s, -NH-); 10.70 (1H, s, -N**H**). ¹³C NMR (75 MHz, DMSO- d_{s} /TMS): δ ppm: 24.25, 26.91 (aliphatic C), 37.34 (S-CH₂), 60.58 (O-CH₂), 115.48, 118.86, 120.80, 127.44, 130.02, 130.27, 130.61, 132.42, 133.05, 133.88, 143.52, 151.93 (Ar-C), 152.30 and 153.49 (triazole C3 and C5), 166.53 and 168.28 (CO, amide), 196.99 (Ar-COCH₂). Anal. calc. for C₂₇H₂₅N₅O₄S C: 62.90, H:4.89, N:13.58, S:6.22; found C:63.00 H:5.07, N:13.10, S:5.81. LR-MS (ESI) (m/z): Calc. for (M+Na)+: 538.16, found: 538.15. Calc. for (M-H): 514.16, found: 514.20.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4phenyl-4*H*-1,2,4-triazol-3-yl)sulfanyl]-*N*-(4-nitro phenyl)acetamide (**18**): M.p. 273.0-274.0 °C (lit. 250 °C, (Yurttaş, Evren, Kubilay, & Temel, 2021)), yield 57%. TLC Rf: 0.35. IR ν (cm⁻¹): 3225, 3161 (N-H str), 3107-2841 (C-H str), 1697, 1667 (C=O str), 1559 (C=N str), 681 (C-S str). ¹H NMR (300 MHz, DMSO-d₆) δ ppm: 1.06 (3H, t, *J*= 6.6 *Hz*, 1/3 mol CH₃CH₂OH); 1.99 (3H, s, -COCH₃); 3.40-3.49 (2H, m, 1/3 mol CH₃CH₂OH); 4.26 (2H, s, -S-CH₂-); 4.37 (1H, t, J = 5.1 Hz, 1/3 mol CH₃CH₂OH); 5.03 (2H, s, -OCH₂); 6.78 (2H, d, J = 9 Hz, Ar-H); 7.41 (2H, d, J = 9 Hz, Ar-H); 7.49-7.59 (5H, m, Ar-H); 7.82 (2H, d, J = 9 Hz, Ar-H); 8.25 (2H, d, J = 9 Hz, Ar-H); 9.79 (1H, s, -NH-); 10.97 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/TMS): δ ppm: 15.52, 24.27 (aliphatic C), 37.78 (S-CH₂), 60.48 (O-CH₂), 115.36, 119.32, 120.94, 125.54, 133.88, 142.86, 145.32, 150.46 (Ar-C), 152.12 and 153.60 (triazole C3 and C5), 167.15 and 168.30 (CO, amide). Anal. calc. for C₂₅H₂₂N₆O₅S.1/3 mol C₂H₅OH C: 57.78 H:4.47, N:15.85, S:6.05; found C:57.35 H:4.65, N:15.56, S:6.20. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 541.14, found: 541.00. Calc. for (M-H)⁻: 517.14, found: 517.00.

2-[(5-{[4-(Acetylamino)phenoxy]methyl}-4phenyl-4H-1,2,4-triazol-3-yl)sulfanyl]-N-(2-methyl-5-nitrophenyl)acetamide (19): M.p. 158.9-160.7 °C, yield 49%. TLC Rf: 0.28. IR v (cm⁻¹): 3258, 3185 (N-H str), 3127-2863 (C-H str), 1688, 1657 (C=O str), 1535 (C=N str), 693 (C-S str). ¹H NMR (300 MHz, DMSO-d₂) δ ppm: 1.06 (3H, t, J= 6.6 Hz, 0.5 mol CH, CH, OH); 1.98 (3H, s, -COCH,); 2.36 (3H, s, Ar-CH₃); 3.40-3.49 (2H, m, 0.5 mol CH₃CH₂OH); 4.26 (2H, s, -S-CH₂-); 4.38 (1H, t, J= 5 Hz, 0.5 mol CH₂CH₂OH); 5.05 (2H, s, -OCH₂); 6.79 (2H, d, J= 8 *Hz*, Ar-**H**); 7.41 (2H, d, *J*= 8 *Hz*, Ar-**H**); 7.50-7.59 (6H, m, Ar-H); 7.95 (1H, dd, *J*= 2.4 Hz and 9 Hz, Ar-H); 8.49 (1H, d, J= 2.4 Hz, Ar-H); 9.80 (1H, s, -NH-); 10.01 (1H, s, -NH-). ¹³C NMR (75 MHz, DMSO- d_{c}/TMS): δ ppm: 18.55, 19.03, 24.25 (aliphatic C), 36.77 (S-CH₂), 60.59 (O-CH₂), 115.48, 118.31, 119.90, 120.80, 127.44, 130.27, 130.62, 131.92, 133.04, 133.89, 137.45, 139.13, 146.18, 152.08 (Ar-C), 152.37 and 153.48 (triazole C3 and C5), 167.00 and 168.27 (CO, amide). Anal. calc. for C₂₆H₂₄N₆O₅S.1/2 mol C₂H₅OH C: 57.93, H:4.86, N:15.01, S:5.54; found C:57.24 H:4.81, N:14.70, S:5.42. LR-MS (ESI) (m/z): Calc. for (M+Na)⁺: 555.15, found: 555.15. Calc. for (M-H): 531.15, found: 531.15.

Ethyl 4-[({[5-{[4-(acetylamino)phenoxy]methyl}-4-phenyl-4*H*-1,2,4-triazol-3-yl]sulfanyl}acetyl) amino]benzoate (**20**): M.p. 251.4-252.0 °C, yield 60%. TLC Rf: 0.31. IR ν (cm⁻¹): 3256, 3185 (N-H str), 3120-**402** 2870 (C-H str), 1711, 1663 (C=O str), 1545 (C=N str), 694 (C-S str). ¹H NMR (300 MHz, DMSO-d_z) δ ppm: 1.31 (3H, t, J= 6 Hz, CH, CH, O-); 1.99 (3H, s, -COCH₂); 2.73 and 2.89 (s, DMF peak); 4.23 (2H, s, -S-CH₂-); 4.29 (2H, q, *J*= 7 *Hz*, CH₂CH₂O-); 5.03 (2H, s, -OCH₂); 6.78 (2H, d, J= 9 Hz, Ar-H); 7.41 (2H, d, J= 9 Hz, Ar-H); 7.49-7.59 (5H, m, Ar-H); 7.71 (2H, d, J= 9 Hz, Ar-H); 7.94 (2H, d, J= 9 Hz, Ar-H); 9.79 (1H, s, -NH-); 10.70 (1H, s, -NH). ¹³C NMR (75 MHz, DMSO-d₆/TMS): δ ppm: 14.68, 24.25 (aliphatic C), 37.31 (S-CH₂), 60.94 (O-CH₂), 115.47, 118.97, 120.79, 124.97, 127.44, 130.27, 130.60, 130.79, 133.05, 133.88, 143.55, 151.92 (Ar-C), 152.29 and 153.49 (triazole C3 and C5), 165.74 (CO, ester), 166.52 and 168.26 (CO, amide). Anal. calc. for $C_{28}H_{27}N_5O_5S$ C: 61.64, H:4.99, N:12.84, S:5.88; found C:62.32, H:5.09, N:13.09, S:5.90. LR-MS (ESI) (m/z): Calc. for (M+Na)+: 568.17, found: 568.15. Calc. for (M-H): 544.17, found: 544.20.

Biological methods

Cell culture

Cell culture studies were conducted using lung cancer (A549), chronic myelogenous leukemia (K562), breast cancer (MCF-7), prostate cancer (PC-3), neuroblastoma (SH-SY5Y), and mouse embryonic fibroblast (NIH/3T3) cells as human cancer cell lines. The cells were grown, and cell passage was carried out according to previously reported methods (Demirbolat et al., 2022).

Cell viability assay

Cell viability was assessed using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Cells were seeded in 96-well plates at a density of 1×10^4 cells per well and incubated overnight. Subsequently, the cells were exposed to varying concentrations (1-50 µM) of the synthesized compounds (**9-20**) for 48 hours. Following the initial incubation period, MTT was introduced to each well at a final concentration of 0.5 mg/mL, and the cells were further incubated for an additional 4 hours. After aspirating the growth medium, 100 µl of SDS buffer was introduced to dissolve the purple formazan product. Subsequently, a microplate reader (BioTek, Winooski, VT, USA) was employed to measure the absorbances at 570 nm and 630 nm wavelengths (Demirbolat et al., 2022).

Preparation of human mPGES-1 membrane fraction and PGES activity assay

Human mPGES-1 membrane fractions were prepared as described before (Demirbolat et al., 2022). The inhibitory potential of the selected compounds was assessed through the 2-thiobarbituric acid (TBA) – malondialdehyde (MDA) assay, following a previously established protocol (Larsson et al., 2019). For reference, 1-{6-chloro-5-methyl-1-[6-(trifluoromethyl)pyridin-2-yl]-1*H*-benzimidazol-2-yl}-*N*-(oxolan-3-yl)piperidine-4-carboxamide (compound **118**), a recognized potent mPGES-1 inhibitor, was employed (Larsson et al., 2019).

In vitro COX-1/2 enzyme inhibition assay

The inhibitory activities of all synthesized compounds on COX-1/2 were assessed using an enzyme immunoassay-based approach, specifically the 'COX (ovine/human) Inhibitor Screening Assay' kit (#560131, Cayman Chemical, Ann Arbor, MI, USA). To ascertain their selectivity for mPGES-1 compared to COX-1/2, a combination of equal amounts of COX-1 and COX-2 enzymes was prepared for this study. Each assay was conducted in triplicate, and the inhibitory efficacy of the tested compounds was quantified as the percentage inhibition against COX-1/2 enzymes at a concentration of 100 μ M. The COX-1/2 determination experiments were carried out according to the method specified in the literature (Demirbolat et al., 2022).

In silico methods

Prediction of ADMET profiles

The solubility properties and structural descriptors of the novel paracetamol-triazole conjugates, compounds **9-20**, were evaluated. Target compounds were assessed for their LogP, molecular weight, solubility, topological polar surface area, count of hydrogen bond acceptors/donors and rotatable bonds, %ABS, etc. Moreover, compounds **9-20** were

examined for Lipinski's Rule of 5 for their compliance (Lipinski, Lombardo, Dominy, & Feeney, 1997). All the data mentioned above were procured from the online web server SwissADME (Daina, Michielin, & Zoete, 2017). In silico toxicological parameters were predicted by OSIRIS Data Warrior software (Sander, Freyss, Von Korff, & Rufener, 2015)which molecular families are present, which structural motifs correlate with measured properties, and which tiny structural changes cause large property changes. Data visualization and analysis software with sufficient chemical intelligence to support chemists in this task is rare. In an attempt to contribute to filling the gap, we released our in-house developed chemistry aware data analysis program DataWarrior for free public use. This paper gives an overview of DataWarrior's functionality and architecture. Exemplarily, a new unsupervised, 2-dimensional scaling algorithm is presented, which employs vectorbased or nonvector-based descriptors to visualize the chemical or pharmacophore space of even large data sets. DataWarrior uses this method to interactively explore chemical space, activity landscapes, and activity (Sander, Freyss, Von Korff, & Rufener, 2015) (http://www.openmolecules.org/datawarrior).

RESULTS AND DISCUSSION Synthesis and characterization

Compounds 1-5 were synthesized starting from paracetamol as described in previously reported methods (Demirbolat et al., 2022). The synthesis of compounds 9-20 followed the reaction protocol as outlined in Scheme 1. 3H-1,2,4-Triazole-3-thione derivatives (6-8) were synthesized through the cyclocondensation process of compounds 3-5. This reaction occurred in the presence of TEA in ethanol to prevent the hydrolysis of the acetamido moiety. N-aryl-2-chloroacetamides Conversely, (**I-IV**) were prepared by reacting substituted anilines with 2-chloroacetylchloride under the influence of TEA. In final step, triazoles 6-8 were treated with compounds I-IV in DMF to yield target compounds 9-20 (Kulabaş et al., 2016).

Kulabaş, Gürboğa, Bingöl Özakpınar, Liu, Jakobsson, Danış, Ogan, Küçükgüzel



Scheme 1. Synthetic route to compounds 9-20. Key to reagents: a. $BrCH_2COOC_2H_5$, K_2CO_3 , acetone; b. NH, NH, H, O, EtOH; c. R, NCS, EtOH; d. TEA/EtOH, reflux; e. I-IV, K_2CO_3 , DMF f. ClCOCH₂Cl, TEA, DCM.

Compound	R ₁	R ₂	Compound	R ₁	R ₂
9	-CH ₃	4-COCH ₃	15	$-C_{2}H_{5}$	2-CH ₃ -5-NO ₂
10	-CH ₃	4-NO ₂	16	$-C_{2}H_{5}$	$4\text{-}\mathrm{COOC}_{2}\mathrm{H}_{5}$
11	-CH ₃	2-CH ₃ -5-NO ₂	17	$-C_{6}H_{5}$	4-COCH ₃
12	-CH ₃	4-COOC ₂ H ₅	18	$-C_{6}H_{5}$	4-NO ₂
13	$-C_{2}H_{5}$	4-COCH ₃	19	$-C_{6}H_{5}$	2-CH ₃ -5-NO ₂
14	$-C_{2}H_{5}$	4-NO ₂	20	-C ₆ H ₅	4-COOC ₂ H ₅

Among the prepared triazoles (6-8), only compound 8 was synthesized by Yurttaş et al. (Yurttaş et al., 2021). Still, the ring-closing reaction was carried out in 2N KOH whereas we used triethylamine in the ethanolic medium. Our study is unique in this respect, and the structures of compounds 6-8 were confirmed with spectral data for compound 8 and other triazole rings. In the IR spectra of triazoles 6-8, the detection of a C=O band at 1679-1652 cm⁻¹ belonging to paracetamol moiety as well as the C=S band at 1275-1240 cm⁻¹ supported the formation of 1,2,4-triazole-3-thione. Moreover, 1,2,4-triazole N-H proton was detected at 13.83-14.06 ppm in a higher energy field than the thiosemicarbazide NH protons (Tatar et al., 2015).

The synthesized compounds **9-20** were assessed for purity using TLC as well as elemental analysis, and their structures were elucidated by FTIR, ¹H NMR, ¹³C-NMR and LC-MS spectral data. The IR spectra of compounds 9-20 exhibited distinctive N-H and C=O stretching bands, appearing at 3258-3159 cm⁻¹ and 1669-1653 cm⁻¹, respectively. Also, the disappearance of the C=S stretching band of the 1,2,4-triazole-3thione and the detection of the C-S-C stretching band at 693-635 cm⁻¹ confirms these structures. In the ¹H NMR spectra of 9-20, peaks at around 9.79-9.84 ppm and 9.99-10.97 ppm were attributed to the N-H protons of paracetamol and thioacetanilide moieties, respectively, and N-H signals of the 1,2,4-triazol-3-thione were not detectable (Kulabaş et al., 2016). Moreover, S-CH₂ protons were observed at 4.16-4.27 ppm as a singlet peak with 2H integration. In the ¹³C NMR spectra of compound 9-20, signals at about 166.52-168.70 ppm as two peaks that were attributed to the paracetamol and thioacetanilide carbonyl peak (C=O) while C3 and C5 carbon of the 1,2,4-triazole

ring were observed between 152.09-153.71 ppm (Demirbolat et al., 2022). On the other hand, the resonances at 196.97-196.99 ppm were attributed to the acetyl C=O for compounds 9, 13, 17 and signals at about 165.73-165.74 ppm were attributed to the ester C=O for compounds 12, 16, 20. Low-resolution mass spectra (LR-MS) confirmed the molecular weights of compounds 9-20 with determined both [M+Na]⁺ and [M-H]⁻ ion peaks. Finally, elemental analysis data obtained for all synthesized compounds demonstrated consistency with their assigned structures. Among the target compounds, compound 18 was previously synthesized by Yurttaş et al., (Yurttaş et al., 2021) in the presence of K₂CO₂ in acetone, and its matrix metalloproteinase-9 (MMP-9) inhibition potential was reported. Our study differs from the other reported study in terms of the difference in the synthesis method and the bioactivity studies performed.

Biological activity studies

Cytotoxic effects of compounds 9-20

Compounds 9-20 were screened for their antiproliferative properties against A549, K562, MCF-7, PC-3, SH-SY5Y, and cytotoxic effects in NIH/3T3 cells using MTT assay. Table 1 provides a comprehensive summary of the outcomes from the initial screening conducted at a concentration of 10 µM. Notably, negligible cell inhibitory effects were observed in the samples treated with DMSO. Among the various cell lines tested, MCF-7 exhibited the most pronounced inhibitory response to the synthesized compounds, yielding inhibition values ranging from 17.29% to 25.06%. It was found that these compounds did not show significant inhibition against all selected cell lines in general. However, the highest inhibition value against healthy cell NIH3T3 was 23.04%, and therefore, they were considered not to be cytotoxic.

Table 1. Cytotoxicity profiles of compounds 9-20 against cancer cell lines and NIH3T3 cell line.

Compound	Lah ID cadaa	Inhibition %						
	Lab ID codes	A549	K562	MCF7	PC-3	SH-SY5Y	NIH3T3	
9	KUC16D590	9.12	-3.86	13.28	-9.44	5.67	0.18	
10	KUC16D577	2.34	-3.72	9.52	-6.92	-5.16	-2.14	
11	KUC16D584	0.58	-6.41	8.02	-12.03	-4.59	-7.32	
12	KUC16D588	6.43	2.33	19.17	-13.11	-4.85	7.32	
13	KUC16D621	3.51	-3.86	12.91	-1.87	1.48	5.89	
14	KUC16D609	10.53	-1.60	17.42	7.93	-9.65	13.57	
15	KUC16D616	4.68	13.63	-2.26	0.50	0.82	-8.39	
16	KUC16D620	9.71	-2.04	18.55	0.72	0.01	10.18	
17	KUC16D683	17.31	4.01	17.29	5.48	3.47	13.04	
18	KUC16D671	15.91	-4.37	21.30	8.93	8.98	21.07	
19	KUC16D678	6.32	9.62	9.52	-0.65	7.50	7.86	
20	KUC16D682	10.76	0.95	25.06	18.37	8.01	23.04	

Inhibition of mPGES-1 and COX-1/2 enzymes

Inhibitory activities of compounds 9-20 were screened towards mPGES-1 and COX-1/2 enzymes. Compound 118 was used as a reference compound for the mPGES-1 enzyme (Larsson et al., 2019). Preparation of a protein mixture containing equal amounts of COX-1 and COX-2 enzymes was performed to evaluate whether the tested compound had the potential to significantly inhibit COX-1 or COX-2. This approach enables the determination of substantial inhibitory activity against the combined COX-1 and COX-2 enzymes. Dexketoprofen, ibuprofen, and celecoxib were used as reference compounds for COXs enzymes. As seen in Table 2, it was observed that the target compounds had a significant effect against neither the mPGES-1 enzyme at 10 μ M dose nor the COX-1/2 enzyme mix at 100 μ M dose.

German		% Inhibition of COX-1/2		
Compound	% Inhibition	P value	P value (-log10)	at 100 μM
9	3.895	0.601	0.221	19.1
10	0.034	0.938	0.028	17.5
11	-1.087	0.495	0.306	16.6
12	2.506	0.435	0.362	14.7
13	1.686	0.341	0.467	20.0
14	-2.152	0.067	1.174	10.8
15	0.294	0.611	0.214	12.2
16	1.912	0.326	0.487	10.6
17	0.203	0.891	0.050	15.8
18	0.072	0.951	0.022	9.1
19	1.363	0.284	0.547	5.8
20	-1.663	0.371	0.431	16.6
Compd 118ª	73.780	0.000	4.030	
Dexketoprofen				65.3
Ibuprofen				68.1
Celecoxib				N.D. ^b

Table 2. Inhibitory properties of compounds 9-20 against mPGES-1 and COX-1/2.

^a Compound 118, reported by Larsson et. al.(Larsson et al., 2019), was used as reference compound.

^bNot determined; inhibition at this concentration is above the linear value of the calibration plot (too high inhibition)

In silico ADMET predictions

SMILES codes of the compounds **9-20** were prepared using the ACD/ChemSketch v. 12.0 to estimate their ADMET parameters through SwissADME software (http://www.swissadme.ch). The range of hydrogen bond acceptors in the tested compounds was either 6 or 7, and the number of rotatable bonds exhibited variability within the scope of 11 to 14. Notably, none of the tested compounds had more than two hydrogen bond donors (Table 3). The association between low aqueous solubility and limited absorption is well-established. Remarkably, the synthesized compounds demonstrated favorable logS values exceeding -5.2.

Furthermore, the estimated intestinal absorption (%ABS) of the synthesized compounds was calculated according to the methodology reported by Zhao et al. (Zhao et al., 2002)diffusion, or perfusion processes can become the rate-limited step. The absorption data of 238 drugs have been classified into either dissolution or diffusion rate-limited based on an equilibrium method developed from solubility, dose, and percentage of absorption. A nonlinear absorption model derived from first-order kinetics has been developed to identify the relationship between percentage of drug absorption and molecular descriptors. Regression analysis was performed between percentage of absorption and molecular descriptors. The descriptors used were ClogP, molecular polar surface area, the number of hydrogen-bonding acceptors and donors, and Abraham descriptors. Good relationships were found between absorption and Abraham descriptors or ClogP. The absorption models can predict the following three BCS (Biopharmaceutics Classification Scheme, and determined between 50.6-60.5% for compounds 9-20. Adhering to Lipinski's rule of five, it is conventionally recommended that the molecular weight remains below 500 Da. However, it is noteworthy that compounds 17-20, characterized by a phenyl ring at the 4th position of the triazole moiety, appear to deviate from this criterion.

Compound	LogP _{o/w}	LogS (ESOL)	nON	nOHN	nRot	TPSA	%ABS	Lipinski Rule nviol
9	2.04	-3.29	6	2	11	140.51	60.5	0
10	1.35	-3.40	7	2	11	169.26	50.6	0
11	1.55	-3.70	7	2	11	169.26	50.6	0
12	2.45	-3.65	7	2	13	149.74	57.3	0
13	2.37	-3.49	6	2	12	140.51	60.5	0
14	1.68	-3.59	7	2	12	169.26	50.6	0
15	1.93	-3.90	7	2	12	169.26	50.6	0
16	2.72	-3.85	7	2	14	149.74	57.3	0
17	3.32	-4.71	6	2	12	140.51	60.5	1
18	2.81	-4.83	7	2	12	169.26	50.6	1
19	3.10	-5.13	7	2	12	169.26	50.6	1
20	3.66	-5.09	7	2	14	149.74	57.3	1

Table 3. Solubility and molecular descriptors of compounds 9-20 from SwissADME.

LogPo/w: Consensus; LogS (ESOL): Estimating aqueous solubility from molecular structure; nON: Number of hydrogen acceptors; nOHN: Number of hydrogen donors; nRot: Number of rotatable bonds; TPSA: Topological polar surface area; %ABS: Percentage of absorption.

SwissADME server was also used to boiled egg plot denotes. In this graphic, the white area is defined as intestinal absorption, while the gray area is poor intestinal absorption. The yellow region, which designates effective brain penetration and favorable intestinal absorption, is demarcated. None of the synthesized compounds exhibited the potential to cross the blood-brain barrier as predicted. All target compounds that exhibit poor intestinal absorption have been found to be potential substrates of P-glycoprotein except compounds 18 and 19. According to the estimated toxicity profiles of the compounds, only compounds 9, 13, and 17 containing the acetyl group showed mutagenic effects, while none of the compounds showed tumorigenic and irritant profiles (Figure 2).

CONCLUSION

In conclusion, a new series of paracetamol-triazole conjugates **9-20** were synthesized, and their structures were characterized by elemental analyses besides spectral methods. Cytotoxic effects of the target compounds were tested on A549, K562, MCF-7, PC-3, SH-SY5Y, and NIH-3T3 cells. Furthermore, a comprehensive evaluation was conducted on compounds **9-20** to assess their inhibitory potential against mPGES-1 and COX-1/2 enzymes. None of these compounds exhibited significant antiproliferative effect and mPGES-1 inhibition, as well as COX-1/2 inhibition, too. However, compounds **9-20** showed no cytotoxic effect against NIH-3T3 cells, concluding that different biological effects of these safe compounds can be evaluated in our future studies.



Figure 2. Graphical distribution of compounds according to the boiled egg predictive model and their toxicity profiles.

ACKNOWLEDGMENTS

This work was supported by The Scientific and Technological Research Council of Turkey (TÜBİTAK - Grant no. 218S549) and the European Union's Horizon 2020 research project ArthritisHeal under the Marie Skłodowska-Curie grant agreement (No. 812890).

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHOR CONTRIBUTION STATEMENT

The hypothesis of this study were developed and literature research was carried out by N.K. and İ.K. Compounds were designed, synthesized and characterized by N.K. and İ.K. The antiproliferative activity of synthesized compounds were determined by M.G. and Ö.B.Ö. Their inhibitory activity against mPGES-1 enzyme were tested by J.L. and P.-J.J. Then inhibition potential of these compounds were evaluated against COX-1/2 enzymes by Ö.D. and A.O. Finally, manuscript preparation and critical reviews were realized by N.K., M.G., Ö.B.Ö., J.L., J.J., Ö.D., A.O. and İ.K.

REFERENCES

Akasaka, H., So, S. P., & Ruan, K. H. (2015). Relationship of the Topological Distances and Activities 408

between mPGES-1 and COX-2 versus COX-1: Implications of the Different Post-Translational Endoplasmic Reticulum Organizations of COX-1 and COX-2. Biochemistry, 54(23), 3707-3715. https://doi.org/10.1021/acs.biochem.5b00339

- Bülbül, B., Ding, K., Zhan, C. G., Çiftçi, G., Yelekçi, K., Gürboğa, M., ... Küçükgüzel, İ. (2022). Novel 1,2,4-triazoles derived from Ibuprofen: synthesis and in vitro evaluation of their mPGES-1 inhibitory and antiproliferative activity. Molecular Diversity, (Early Access). https://doi.org/10.1007/ s11030-022-10551-0
- Chang, H. H., Song, Z., Wisner, L., Tripp, T., Gokhale, V., & Meuillet, E. J. (2012). Identification of a novel class of anti-inflammatory compounds with anti-tumor activity in colorectal and lung cancers. Investigational New Drugs, 30, 1865–1877.
- Crofford, L. J. (1997). COX-1 and COX-2 tissue expression: implications and predictions. The Journal of Rheumatology. Supplement, 49, 15-19. Retrieved from http://europepmc.org/abstract/ MED/9249646
- Daina, A., Michielin, O., & Zoete, V. (2017). SwissADME: A free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. Scientific Reports, 7, 1-13. https://doi.org/10.1038/ srep42717

- Demirbolat, İ., Kulabaş, N., Gürboğa, M., Bingöl-Özakpınar, Ö., Çiftçi, G., Yelekçi, K., ... Küçükgüzel, İ. (2022). Synthesis and evaluation of antiproliferative and mPGES-1 inhibitory activities of novel carvacrol-triazole conjugates. Organic Communications, 15(4), 356–377.
- Demirci, A., Karayel, K. G., Tatar, E., Okullu, S. Ö., Unübol, N., Taşli, P. N., ... Küçükgüzel, I. (2018). Synthesis and evaluation of novel 1,3,4-thiadiazole-fluoroquinolone hybrids as antibacterial, antituberculosis, and anticancer agents. *Turkish Journal of Chemistry*, 42(3), 839– 858. https://doi.org/10.3906/kim-1710-35
- Erensoy, G., Ding, K., Zhan, C. G., Çiftçi, G., Yelekçi, K., Duracık, M., ... Küçükgüzel, İ. (2023).
 Synthesis, in vitro and in silico studies on novel 3-aryloxymethyl-5-[(2-oxo-2-arylethyl)sulfanyl]-1,2,4-triazoles and their oxime derivatives as potent inhibitors of mPGES-1. *Journal of Molecular Structure*, 1272, 134154. https://doi.org/10.1016/j.molstruc.2022.134154
- Graham, G. G., Davies, M. J., Day, R. O., Mohamudally, A., & Scott, K. F. (2013). The modern pharmacology of paracetamol: therapeutic actions, mechanism of action, metabolism, toxicity and recent pharmacological findings. 201–232. https://doi. org/10.1007/s10787-013-0172-x
- Haider, B., & Hamada, M. I. (2012). 4-Thiazolidinones in Heterocyclic Synthesis: Synthesis of Novel Enaminones, Azolopyrimidines and 2-Arylimino-5-arylidene-4-thiazolidinones. *Molecules*, 17, 6362–6385. https://doi.org/10.3390/ molecules17066362
- Hanaka, H., Pawelzik, S. C., Johnsen, J. I., Rakonjac, M., Terawaki, K., Rasmuson, A., ... Rådmark, O. (2009). Microsomal prostaglandin E synthase 1 determines tumor growth in vivo of prostate and lung cancer cells. *Proceedings of the National Academy of Sciences*, 106, 18757–18762.
- He, S., Li, C., Liu, Y., & Lai, L. (2013). Discovery of highly potent microsomal prostaglandin E2 synthase 1 inhibitors using the active conformation structural model and virtual screen. *Journal of Medicinal Chemistry*, 56(8), 3296–3309. https://

doi.org/10.1021/jm301900x

- Howe, L. R., Subbaramaiah, K., Kent, C. V., Zhou, X. K., Chang, S. H., Hla, T., ... Dannenberg, A. J. (2013). Genetic deletion of microsomal prostaglandine synthase-1 suppresses mouse mammary tumor growth and angiogenesis. *Prostaglandins & Other Lipid Mediators*, 106, 99–105.
- Kock, A., Larsson, K., Bergqvist, F., Eissler, N., Elfman,
 L. H. M., Raouf, J., ... Kogner, P. (2018). Inhibition
 of Microsomal Prostaglandin E Synthase-1
 in Cancer-Associated Fibroblasts Suppresses
 Neuroblastoma Tumor Growth. *EBioMedicine*, 32, 84–92.
- Küçükgüzel, İ, Küçükgüzel, S. G., Rollas, S., & Kiraz, M. (2001). Some 3-thioxo/alkylthio-1,2,4triazoles with a substituted thiourea moiety as possible antimycobacterials. *Bioorganic & Medicinal Chemistry Letters*, 11(13), 1703–1707. Retrieved from http://www.ncbi.nlm.nih.gov/ pubmed/11425542
- Küçükgüzel, İlkay, Tatar, E., Küçükgüzel, Ş. G., Rollas, S., & De Clercq, E. (2008). Synthesis of some novel thiourea derivatives obtained from 5-[(4-aminophenoxy)methyl]-4-alkyl/ aryl-2,4-dihydro-3H-1,2,4-triazole-3-thiones and evaluation as antiviral/anti-HIV and anti-tuberculosis agents. *European Journal of Medicinal Chemistry*, 43(2), 381–392. https://doi. org/10.1016/j.ejmech.2007.04.010
- Küçükgüzel, Ş. G., Küçükgüzel, I., Tatar, E., Rollas, S., Sahin, F., Güllüce, M., ... Kabasakal, L. (2007). Synthesis of some novel heterocyclic compounds derived from diflunisal hydrazide as potential antiinfective and anti-inflammatory agents. *European Journal of Medicinal Chemistry*, 42(7), 893–901. https://doi.org/10.1016/j.ejmech.2006.12.038
- Kulabaş, N., Tatar, E., Bingöl Özakpınar, Ö., Özsavcı, D., Pannecouque, C., De Clercq, E., & Küçükgüzel, İ. (2016). Synthesis and antiproliferative evaluation of novel 2-(4H-1,2,4-triazole-3-ylthio)acetamide derivatives as inducers of apoptosis in cancer cells. *European Journal of Medicinal Chemistry*, *121*, 58– 70. https://doi.org/10.1016/j.ejmech.2016.05.017

- Kulabaş, N., Türe, A., Bozdeveci, A., Krishna, V. S., Alpay Karaoğlu, Ş., Sriram, D., & Küçükgüzel, İ. (2022). Novel fluoroquinolones containing 2-arylamino-2-oxoethyl fragment: Design, synthesis, evaluation of antibacterial and antituberculosis activities and molecular modeling studies. *Journal of Heterocyclic Chemistry*, 59(5), 909–926. https://doi.org/10.1002/jhet.4430
- Larsson, K., Steinmetz, J., Bergqvist, F., Arefin, S., Spahiu, L., Wannberg, J., ... Jakobsson, P. J. (2019).
 Biological characterization of new inhibitors of microsomal PGE synthase-1 in preclinical models of inflammation and vascular tone. *British Journal* of Pharmacology, 176(24), 4625–4638.
- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney,
 P. J. (1997). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, 23, 3–25.
- Morar, C., Cost, L., Lameiras, P., Antheaume, C., & Darabantu, M. (2015). Convergent Versus Divergent Three-Step Synthesis of the First (4-Aminophenoxy)alkanoic Acid–Based Tripodal Melamines. Synthetic Communications, 45(14), 1688–1695. https://doi.org/10.1080/00397911.20 15.1041048
- Murakami, M. (2011). Lipid Mediators in Life Science. Experimental Animals, 60(May 2010), 7–20. https://doi.org/10.1538/expanim.60.7
- Nargund, L. V. G., Reddy, G. R. N., & Hariprasad, V. (1994). Anti-inflammatory Activity of Substituted 1,3,4-Oxadiazoles. *Journal of Pharmaceutical Sciences*, 83(2), 246–248. https://doi.org/10.1002/ jps.2600830226
- Sander, T., Freyss, J., Von Korff, M., & Rufener, C. (2015). DataWarrior: An open-source program for chemistry aware data visualization and analysis. *Journal of Chemical Information and Modeling*, 55(2), 460–473. https://doi.org/10.1021/ci500588j
- Sasaki, Y., Nakatani, Y., & Hara, S. (2015). Role of microsomal prostaglandin E synthase-1 (mPGES-1)-derived prostaglandin E2 in colon carcinogenesis. *Prostaglandins & Other Lipid Mediators*, 121, 42–45.

- Shay, A. E., Diwakar, B. T., Guan, B.-J., Narayan, V., Urban, J. F. J., & Prabhu, K. S. (2017). IL-4 up-regulates cyclooxygenase-1 expression in macrophages. *The Journal of Biological Chemistry*, 292(35), 14544–14555. https://doi.org/10.1074/ jbc.M117.785014
- Svetkin, Y. V., Minlibaeva, A. N., & Khafizova, N. A. (1961). Reaction of ketene with nitrogenous bases. IX. Chloroacetylation of primary aromatic amines. *Zhurnal Obshchei Khimii*, 31, 2023–2023.
- WHO. (2022). Cancer. Retrieved August 30, 2023, from https://www.who.int/news-room/factsheets/detail/cancer
- Yurttaş, L., Evren, A. E., Kubilay, A., & Temel, H. E. (2021). Synthesis of New 1,2,4-Triazole Derivatives and Investigation of Their Matrix Metalloproteinase-9(MMP-9)InhibitionProperties. 59(2), 0–2. https://doi.org/10.23893/1307-2080. APS.05913
- Zengin, M., Unsal Tan, O., Arafa, R. K., & Balkan, A. (2022). Design and synthesis of new 2-oxoquinoxalinyl-1,2,4-triazoles as antitumor VEGFR-2 inhibitors. *Bioorganic Chemistry*, 121, 105696.
- Zhai, W., Hu, Y., Zhang, Y., Zhang, G., Chen, H., Tan, X., ... Wu, J. (2022). A systematic review of phytochemicals from Chinese herbal medicines for non-coding RNAs-mediated cancer prevention and treatment: From molecular mechanisms to potential clinical applications. *Medicine in Novel Technology and Devices*, 16(November). https:// doi.org/10.1016/j.medntd.2022.100192
- Zhao, Y. H., Abraham, M. H., Le, J., Hersey, A., Luscombe, C. N., Beck, G., ... Cooper, I. (2002). Rate-limited steps of human oral absorption and QSAR studies. *Pharmaceutical Research*, 19(10), 1446–1457. https://doi. org/10.1023/A:1020444330011
- Zhaowen, L., Li, Z., Chunfen, X., Yong, Y., Fanbo,
 Z., & Kaixun, H. (2007). Anticancer activities of some arylcarbamoylalkyltriphenylphosphonium chlorides. *Medicinal Chemistry Research*, 16(7–9), 380–391. https://doi.org/10.1007/s00044-007-9050-7