

e-ISSN : 2757-6744 doi : 10.52037/eads.2023.0025

Article Received/Accepted : June, 9 2023 / December, 18 2023 Ethical Committee No : 9/1,08.01.2011

ORIGINAL RESEARCH ARTICLE

The Effect of Body Mass Index on the KYN/TRP Pathway in the Pathogenesis of Periodontitis

Zeliha Güney ¹ and Sema Merve Altıngöz ^{2, *}

¹Department of Periodontology, Faculty of Dentistry, Ankara Medipol University, Ankara, Turkey and ²Department of Periodontology, Faculty of Dentistry, Lokman Hekim University, Ankara, Turkey

*Corresponding Author; smerve.unal@yahoo.com

Abstract

Purpose: The tryptophan-kynurenine (TRP-KYN) pathway is associated with inflammation, and kynurenine pathway (KP) dysregulation is present in overweight and obesity. Meanwhile, obesity and periodontitis are two of the most frequent noncommunicable illnesses, and epidemiological studies show that obesity has a role in the initiation and progression of periodontitis. However, the association between elevated body mass index and KP on periodontal disease etiology is unknown. As a result, our study aims to investigate the possible relationship between TRP/KYN ratio and body mass index (BMI) relationship in periodontitis.

Materials and Methods: The study comprised 20 periodontitis patients (P, Generalized Stage III Grade B, n=20) and 20 healthy persons (C, n=20). Clinical parameters (Bleeding index on probing (BOP), clinical attachment loss (CAL) and pocket depth (PD)), and BMI were recorded at the beginning of the study. Salivary and serum KYN/TRP ratios were analyzed using mass spectrometry—liquid chromatography (LC–MS/MS).

Results: Clinical periodontal parameters were statistically significantly higher in P group than in C group (p<0.05). While there was no change in serum KYN/TRP ratio across groups, salivary KYN/TRP ratio decreased in Group P compared with Group C (p<0.05). The salivary KYN/TRP ratio was positively correlated with the serum KYN/TRP ratio. At the same time, it showed a strong negative correlation with BOP, a moderate correlation with PD and CAL, and lower negative correlation with BMI. **Conclusions:** KP dysregulation due to obesity may increase the risk of developing periodontal disease.

Key words: Periodontitis; Obesity; BMI; KYN/TRP; Tryptophan; Kynurenine

Introduction

Tryptophan, an essential amino acid for human metabolism, is involved in the biosynthesis of significant substances such as neurotransmitters, neurohormones, and niacin.^{1,2} 90% of the metabolism of tryptophan is carried out via the oxidation process, commonly known as the kynurenine pathway (KP) (Figure 1).³ Tryptophan, which differs in tissue and cell localization and substrate specificities, is converted to N-formyl kynurenine by enzymes called tryptophan 2,3-dioxygenase (TDO) and indolamine 2,3-dioxygenase (IDO).⁴ While IDO acts locally and modulates tryptophan levels in response to inflammation, TDO acts systemically by regulating blood tryptophan levels.⁵ Other catabolic pathway enzymes contribute to generate the metabolites of this pathway, which are biologically active and affect cellular activities in physiological and pathological ways.⁴ Under physiological conditions, KP is tightly regulated but changes when the immune system is active. KP dysregulation has been associated with several diseases, including cardiovascular diseases, ^{6–8} depression, ^{9,10} inflammatory bowel disease, ¹¹ diabetes, ¹² schizophrenia, ¹³ neurodegenerative disorders, ¹⁴ cancer ^{15,16} and multiple sclerosis. ⁶

Obesity and periodontitis are two of the most prevalent noncommunicable illnesses, and epidemiological studies show that obesity has a role in periodontitis initiation and progression. ¹⁷ Obesity is recognized to have a chronic low-grade inflammatory state, which is also a prevalent factor in obesity-related diseases. ^{18–21} Furthermore, metabolites such as amino acids are variables that interact with processes involved in metabolic balance. ^{19,21,22} The catabolic pathways of tryptophan appear to be particularly important since they are controlled by dietary and inflammatory signals and are connected to metabolic control and management of calorie intake. ^{23–25} Under normal physiological conditions, TDO performs over 90% of tryptophan metabolism in the liver, but KP may also be activated extrahepatically by IDO driven by inflammatory signals associated with obesity. ^{23,26–31} The extrahepatic IDO activity is commonly expressed or reflected by the KYN/TRP ratio. ²⁶





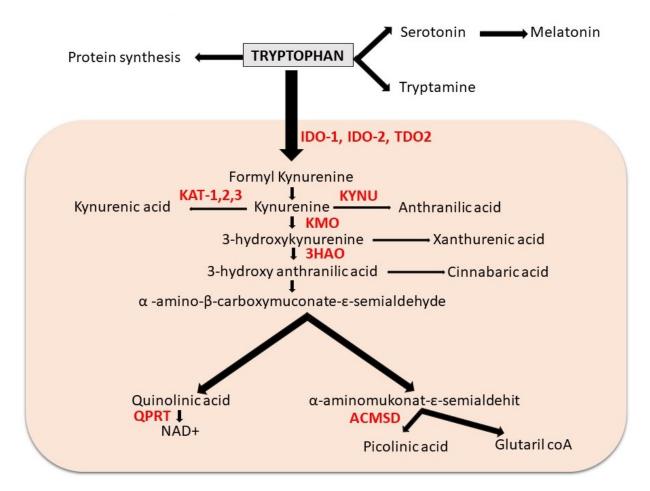


Figure 1. Tryptophan pathway. IDO: indolamine 2,3-dioxygenase; KAT: kynurenine aminotransferase; KYNU: kynureninase; 3-HAO: 3-hydroxy anthranilate 3,4-dioxygenase; QPRT: quinolinic-acid phosphoribosyl transferase; ACMSD: 2-amino-3-carboxymuconate semialdehyde carboxylase.

According to a few studies, KP is additionally related to periodontitis. ^{9,10,32} However, there is no study in the literature investigating the effect of increased KP activation associated with BMI and the impact of this interaction on the risk of periodontitis development. In the light of all these literature findings, our study hypothesizes that as the BMI increases, KP activation will increase, increasing the risk of periodontal disease development. Our study aims to evaluate the salivary and serum kynurenine/tryptophan (KYN/TRP) ratio change between periodontal inflammation.

Material and Methods

Study Design

The study included twenty periodontitis patients (Group P, Stage III, Grade B, n=20; 11 females, 9 males; mean age: 41.56 ±10.56 years) and twenty periodontally healthy individuals (Group C, n=20; 8 females, 12 males; mean age: 39.56±6.51 years) who applied to Ankara University Faculty of Dentistry, Department of Periodontology. The Helsinki Declaration conducted the study, which was approved by the Ankara University Faculty of Dentistry Human Research Ethics Committee (No: 9/1, on 08.01.2011). All individuals who agreed to participate in the study signed a valid informed consent form. The periodontal diagnosis was based on the World Workshop on Periodontal and Peri-Implant Diseases and Conditions (2017). ³³ At stage III, periodontitis has significantly damaged the attachment apparatus, and tooth loss may occur without advanced treatment. The stage is characterized by deep periodontal lesions extending to the middle portion of the root and whose management is com-

plicated by the presence of deep intrabony defects, furcation involvement, and history of periodontal tooth loss/exfoliation. Periodontitis may progress with different rates in individuals, respond less predictably to treatment in some patients, and may or may not influence general health or systemic disease. Grade B shows a moderate rate of progression, and destruction is commensurate with biofilm deposits.³³ Periodontitis was diagnosed in patients with at least 16 natural teeth (excluding third molars) who exhibited interdental radiographic bone loss ≥ 2 in non-adjacent teeth and probing depth (PD)>3 mm in at least two teeth.

Periodontitis Stage III, Grade B was diagnosed using the following conditions:

- Radiographic evidence of alveolar bone loss extending to middle third of the root and beyond.
- Number of tooth loss due to periodontitis ≤ 4
- Teeth involved \geq %30
- ≥5 mm interdental clinical attachment loss (CAL)
- 0.25<% bone loss/age <1.

Individuals without a symptom or no history of periodontal disease, clinically periodontal health (bleeding on probing (BOP) <10%, PD <3 mm), and well-maintained dental hygiene were designated as controls. An expert periodontist (SMA) performed all measurements from six sides of each tooth. Pregnant and lactating mothers, obese individuals, smokers, patients with systemic disorders including diabetes, cardiovascular disease, and cancer, antibiotics or anti-inflammatory drug usage, or medication with calcium channel blockers, phenytoin, or cyclosporine, or patients who had received periodontal therapy during the previous three months were all excluded.

Table 1. Comparison of Demographic and Clinical Periodontal Parameters Between the Groups

Clinical	Control	Periodontitis (P; n=20)	
Parameters	(C; n=20)		
Age (year)	38.00	40.50	
Age (year)	(35.75-40.5)	(34.75-48.50)	
Gender (n, f/m)	12/8	9/11	
PD (mm)	1.59	2.83	
PD (IIIII)	(1.44-1.64)	(2.42-2.96)*	
BOP (%)	5.30	42.00	
DOP (70)	(3.70-1.50)	(31.38-60.45)*	
CAL (mm)	0.00	3.02	
CAL (IIIII)	(0.00-0.00)	(2.83-3.37)*	
BMI	21.20	26.10	
DIVII	(20.63-23.30)	(21.78-28.46)*	

PD: probing depth; BOP: bleeding on probing; CAL: Clinical attachment lost; BMI: body mass index Data as median-interquartile range. Mann-Whitney U test was used. * Statistically significant difference (p<0.05)

Saliva and Serum Samples

Unstimulated saliva was obtained after a 12-hour fasting period in the early hours (9:00 am to 11:00 am). Whole saliva (approximately 2 ml) was collected using sterile polypropylene tubes. The samples were centrifuged at 2800 ×g for 10 minutes.³⁴

Venous blood samples were collected from antecubital vein by venipuncture. The samples were centrifuged at 4000 ×g for 10 minutes. The saliva and blood samples were kept at -80°C until analysis.³⁴

Analysis of KYN/TRP

The salivary and serum KYN and TRP levels were determined using mass spectrometry–liquid chromatography (LC-MS/MS, ESI Source, Thermo Scientific Accessmax), a method modified from Di Gangi et al.³⁵

Statistical Analysis

Statistical analyses were performed using the SPSS 22.0 (SPSS v.22, IBM SPSS Inc., Chicago, IL, USA) program. Shapiro Wilk test was used to evaluate the normal distribution. Mann-Whitney-U test was used for intergroup comparisons of independent variables, whereas Student-t test was used for dependent variables. Spearman correlation was performed to test the relationship between groups. All tests have been conducted with a significance level of α =0.05.

Results

Clinical Parameters

Table 1 shows demographic and clinical periodontal characteristics. The clinical periodontal parameters in P group were considerably higher than C group (p 0.05). Regarding BMI, age, and gender, there was no significant difference between groups (p=0.997, p=0.706, and p=0.100, respectively).

Biochemical Parameters

Table 2 presents the serum and salivary KYN/TRP ratio. When the KYN/TRP ratios between the two groups were compared, only the salivary levels were different, and the C group was statistically significantly higher than the P group (p<0.001).

Table 2. Saliva and serum KYN/TRP ratio in periodontitis and control groups

Control	Periodontitis	
(C; n=20)	(P; n=20)	
0.05	0.02	
(0.03-0.14)*	(0.02-0.03)	
0.04	0.04	
(0.03-0.04)	(0.04-0.05)	
	(C; n=20) 0.05 (0.03-0.14)* 0.04	

Data are expressed as median-interquartile range. Mann Whitney U Test. * Statistically significant difference (p<0.05)

Correlation of Periodontal and Biochemical Parameters

The association coefficients between periodontal clinical markers and salivary and serum KYN/TRP ratios are shown in Table 3. Serum KYN/TRP ratio positively correlated with BOP and BMI (p<0.05). Salivary KYN/TRP ratio was negatively correlated with BMI and clinical periodontal parameters (p<0.05). Serum and salivary KYN/TRP ratios had a significantly negative correlation (p<0.05).

Discussion

Our study aimed to evaluate the change in BMI and KP activity in periodontitis. The salivary KYN/TRP ratio was statistically significantly higher in C group compared to P group, consistent with our hypothesis, and KP activity decreased as the BMI index increased.

The KP pathway is the primary TRP metabolism pathway. Inflammation or immunological activation of the IDO enzyme causes the extrahepatic KP to activate, which causes the accumulation of TRP metabolic end products.^{36 1,37} As far as we know, no published research has investigated the connection among BMI and KP in periodontal inflammation. We compared the salivary and serum KYN/TRP ratio, which indicates IDO activation, ²⁶ between age, gender, and BMI-matched stage III periodontitis patients and periodontally healthy people based on this information.

The enzyme IDO regulates immunity and inflammation by degrading TRP into KYN and other metabolites (Figure 2).³⁸ A decrease in TRP via IDO-activated Tregs and suppressed Th17 cells has previously been demonstrated to reduce proinflammatory responses in various inflammatory disorders.^{39,40} Several cell types have anti-inflammatory properties through inducing IDO expression.⁴¹ As a result, IDO plays a vital role in controlling inflammation in specific contexts, either by preventing inflammation and maintaining the suppressive phenotype of Tregs when IDO is active or by permitting unregulated inflammation and Treg reprogramming when IDO is absent. As a result, our study reveals that the KYN/TRP ratio decreased with periodontal inflammation. This can be interpreted as a decrease in IDO activity and an increase in the expression of proinflammatory cytokines. Consistent with our results, Hao et al. also showed that P.gingivalis, which is a crucial periodontopathogen, reduces IDO expression in gingival samples.⁴² Indeed, Palm et al. demonstrated that P. gingivalis and its gingipains inhibited IDO expression in human gingival fibroblasts, decreasing the host response and possibly further developing P. gingivalis pathogenicity against host immunity.⁴³ Considering the immune nature of periodontal disease, reduction of IDO expression and/or activity, an immunomodulatory mechanism, may serve host-induced periodontal tissue destruction.

The blood concentration of TRP declines in diseases like HIV infection, neurological disorders, and cancer whereas KYN and other TRP catabolites concentrations rise.^{44,45} However, no change in serum KYN/TRP ratio was identified in our investigation. In contrast to our findings, Kurgan et al.³² reported that the serum KYN/TRP ratio was higher in control group than in periodontitis group and no change in the salivary KYN/TRP ratio between groups. They stated that a large amount of TRP is released into the salivary

-		-	-				
Variables	Sex	BMI	PD	BOP	CAL	KYN/TRP ratio (serum)	KYN/TRP ratio (saliva)
KYN/TRP ratio (serum)							-0,364*
Age	0,333*	0,278	-0,019	0,114	0,142	0,257	-0,099
Sex		0,297	-0,028	0,154	-0,020	0,232	-0,262
BMI			0,170	0,247	0,133	0,418*	-0,399*
PD				0,858**	0,886**	0,297	-0,515*
BOP					0,783**	0,363*	-0,605**
CAL						0,299	-0,443*

Table 3. Correlation of biomarkers between clinical periodontal parameters

BMI: Body mass index; PD: probing depth; BOP: bleeding on probing; CAL: Clinical attachment lost; KYN: Kynurenine; TRP: Tryptophan. Values in bold are different from 0 with a significance level alpha <0.05 (Spearmen correlation test *p<0.05; **p<0.001) r = 0.20-0.40 positive and low correlation. r = 0.40-0.60 positive and mild correlation. r = 0.60-0.80 positive and high correlation.

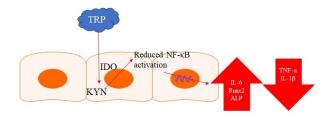


Figure 2. Anti-inflammatory effect of KYN/TRP pathway. TRP: tryptophan; KYN: kynurenine; IDO: indolamine 2,3-dioxygenase; NF- κ B: nuclear factor kappa B; IL-6: interleukin 6, Runx2: Runt related factor 2; ALP: alkaline phosphatase; TNF- α : Tumor necrosis factor alpha; IL-1 β : interleukin 1 beta.

due to enhanced vascularization and protein breakdown in periodontal inflammation. An increase in blood capillaries in gingival connective tissue produced by periodontal inflammation, as well as a rise in gingival crevicular fluid, might explain increased salivary TRP levels. In addition, the inflammation process damages the protein structures in the periodontium, which raises the number of amino acids in saliva. A similar circumstance existed in our study and may have contributed to the lower KYN/TRP ratio.

Obesity is a chronic condition that impairs both general and oral health. ^{46,47} The physiological and molecular processes that may explain the link between obesity and periodontitis are systemic inflammatory alterations in proinflammatory cytokines, hormones, and oxidative stress marker levels. These changes may increase susceptibility to infections and chronic inflammatory diseases. ^{48–50} Numerous epidemiological studies have demonstrated the connection between periodontal disease and obesity. Obese people had a higher odds-ratio for periodontitis than non-obese people, according to a cross-sectional analysis.^{51,52} However, in our study, no statistically significant difference was found in BMI values across the groups; in the P group, BMI was higher than in the C group. Compared with the literature, the salivary KYN/TRP ratio is lower in the P group than in the C group. While the KYN/TRP ratio in the vascular circulation decreases due to low-grade systemic inflammation due to periodontitis, the increased BMI index, which also causes systemic inflammation, may cause an increase in the KYN/TRP ratio. Ultimately, the cumulative effect of these two diseases may have prevented a visible change in the KYN/TRP ratio. Because it is known that obesity increases the serum KYN/TRP ratio⁵³ and causes low-grade chronic inflammation.²⁰ Therefore, in periodontitis where the local inflammatory response is active, the ratio of KYN/TRP in saliva, which contains gingival crevicular fluid and serum, decreases in parallel with the inflammatory response. In addition, it can be interpreted as the increased dietary intake of TRP and the decrease in IDO activity due to increased BMI, resulting in no difference between the groups.

Our study is essential in showing the possible interaction between increased BMI and periodontitis and the effects of these diseases on systemic health. However, the main limitation of our study is that there was no difference between the groups in terms of BMI. Clinical prospective investigations need to be conducted in which groups with different BMI are created and obese individuals are included. Further studies will be precious in showing the effect of possible TRP intake on oral and systemic health, its effect on periodontal disease on systemic health, and the interaction between obesity and periodontitis.

Conclusion

As a result of our study, in which we aimed to evaluate the effects of periodontal inflammation and obesity on KP activity, it can be concluded that IDO activity decreased in the P group and is compatible with host-induced tissue destruction. Although the increase in BMI index reduces IDO activity, the increase in the amount of TRP taken with the diet will also increase the metabolism of this protein. In this context, although IDO activity, which acts locally, decreases with obesity, it can be thought that KP continues its proinflammatory activity through other possible metabolic pathways. However, further studies are needed to evaluate the intergroup variation in obese and non-obese P and C groups.

Author Contributions

All authors have made substantial contributions to conception and design of the study. ZG and SMA have been involved in data collection, data analysis and interpretation, drafting the manuscript and revising it critically and have given final approval of the version to be published.

Conflict of Interest

The authors declare that they have no conflicts of interest.

Authors' ORCID(s)

Z.G. 0000-0001-6897-0773 S.M.A. 0000-0002-9709-6226

References

- Schröcksnadel K, Wirleitner B, Winkler C, Fuchs D. Monitoring tryptophan metabolism in chronic immune activation. Clin Chim Acta. 2006;364(1-2):82–90. doi:10.1016/j.canlet.2004.10.033.
- Widner B, Laich A, Sperner-Unterweger B, Ledochowski M, Fuchs D. Neopterin production, tryptophan degradation, and mental depression—what is the link? Brain Behav Immun. 2002;16(5):590–595.

- 3. Leklem JE. Quantitative aspects of tryptophan metabolism in humans and other species: a review. Am J Clin. 1971;24(6):659–672. doi:10.1093/ajcn/24.6.659.
- Chen Y, Guillemin GJ. Kynurenine pathway metabolites in humans: disease and healthy states. IJTR. 2009;2:IJTR. S2097. doi:10.4137/ijtr.s2097.
- 5. Van der Goot AT, Nollen EA. Tryptophan metabolism: entering the field of aging and age-related pathologies. Trends Mol Med. 2013;19(6):336–344. doi:10.1016/j.molmed.2013.02.007.
- Mangge H, Stelzer I, Reininghaus E, Weghuber D, Postolache TT, Fuchs D. Disturbed tryptophan metabolism in cardiovascular disease. Curr Med Chem. 2014;21(17):1931–1937. doi:10.2174/0929867321666140304105526.
- Wang Q, Zhang M, Ding Y, Wang Q, Zhang W, Song P, et al. Activation of NAD (P) H oxidase by tryptophanderived 3-hydroxykynurenine accelerates endothelial apoptosis and dysfunction in vivo. Circ Res. 2014;114(3):480–492. doi:10.1161/CIRCRESAHA.114.302113.
- Wang Y, Liu H, McKenzie G, Witting PK, Stasch JP, Hahn M, et al. Kynurenine is an endothelium-derived relaxing factor produced during inflammation. Nat Med. 2010;16(3):279–285.
- 9. Moon J, Cheong N, Yang S, Kim I, Chung H, Jeong Y, et al. Lipopolysaccharide-induced indoleamine 2, 3-dioxygenase expression in the periodontal ligament. J Periodontal Res. 2013;48(6):733-739. doi:10.1111/jre.12063.
- Nisapakultorn K, Makrudthong J, Sa-Ard-Iam N, Rerkyen P, Mahanonda R, Takikawa O. Indoleamine 2, 3-dioxygenase expression and regulation in chronic periodontitis. J Periodontol. 2009;80(1):114–121. doi:10.1902/jop.2009.080315.
- Jones SP, Guillemin GJ, Brew BJ. The kynurenine pathway in stem cell biology. IJTR. 2013;6:IJTR. S12626.
- Monasterio G, Budini V, Fernández B, Castillo F, Rojas C, Alvarez C, et al. IL-22–expressing CD 4+ AhR+ T lymphocytes are associated with RANKL-mediated alveolar bone resorption during experimental periodontitis. J Periodontal Res. 2019;54(5):513– 524.
- Mondanelli G, Coletti A, Greco FA, Pallotta MT, Orabona C, Iacono A, et al. Positive allosteric modulation of indoleamine
 3-dioxygenase 1 restrains neuroinflammation. PNAS. 2020;117(7):3848–3857. doi:10.1073/pnas.1918215117.
- Lovelace MD, Varney B, Sundaram G, Franco NF, Ng ML, Pai S, et al. Current evidence for a role of the kynurenine pathway of tryptophan metabolism in multiple sclerosis. Front immunol. 2016;7:246. doi:10.3389/fimmu.2016.00246.
- Cihan M, Dogan O, Ceran Serdar C, Altuncekic Yildirim A, Kurt C, Serdar MA. Kynurenine pathway in Coronavirus disease (COVID-19): potential role in prognosis. J Clin Lab Anal. 2022;36(3):e24257. doi:10.1002/jcla.24257.
- Guillemin GJ. Quinolinic acid, the inescapable neurotoxin. FEBS J. 2012;279(8):1356–1365. doi:10.1111/j.1742-4658.2012.08485.x.
- Arboleda S, Vargas M, Losada S, Pinto A. Review of obesity and periodontitis: an epidemiological view. Br Dent J. 2019;227(3):235–239. doi:10.1038/s41415-019-0611-1.
- Blüher M. Adipose tissue dysfunction in obesity. Exp Clin Endocrinol Diabetes. 2009:241–250. doi:10.1055/s-0029-1192044.
- 19. De Heredia FP, Gómez-Martínez S, Marcos A. Obesity, inflammation and the immune system. Proc Nutr Soc. 2012;71(2):332– 338. doi:10.1017/S0029665112000092.
- 20. Hotamisligil GS. Inflammation and metabolic disorders. Nature. 2006;444(7121):860–867. doi:10.1038/nature05485.
- Maury E, Brichard S. Adipokine dysregulation, adipose tissue inflammation and metabolic syndrome. Mol Cell Endocrinol. 2010;314(1):1–16. doi:10.1016/j.mce.2009.07.031.
- 22. Lischka J, Schanzer A, Hojreh A, Ba Ssalamah A, Item CB, de Gier C, et al. A branched-chain amino acid-based metabolic

score can predict liver fat in children and adolescents with severe obesity. Pediatr Obes. 2021;16(4):e12739.

- 23. Bernard A, Le May C, Dastugue A, Ayer A, Blanchard C, Martin JC, et al. The tryptophan/kynurenine pathway: A novel crosstalk between nutritional obesity, bariatric surgery and taste of fat. Nutrients. 2021;13(4):1366. doi:10.3390/nu13041366.
- 24. Liu JJ, Movassat J, Portha B. Emerging role for kynurenines in metabolic pathologies. Curr Opin Clin Nutr Metab Care. 2019;22(1):82–90. doi:10.1097/MC0.00000000000529.
- Stone TW, McPherson M, Darlington LG. Obesity and cancer: existing and new hypotheses for a causal connection. EBioMedicine. 2018;30:14–28. doi:10.1016/j.ebiom.2018.02.022.
- Badawy AA, Guillemin G. The plasma [kynurenine]/[tryptophan] ratio and indoleamine 2, 3-dioxygenase: time for appraisal. IJTR. 2019;12:1178646919868978. doi:10.1177/1178646919868978.
- 27. Chaves Filho AJM, Lima CNC, Vasconcelos SMM, de Lucena DF, Maes M, Macedo D. IDO chronic immune activation and tryptophan metabolic pathway: A potential pathophysiological link between depression and obesity. Prog Neuropsychopharmacol Biol Psychiatry. 2018;80:234–249.
- 28. Dounay AB, Tuttle JB, Verhoest PR. Challenges and opportunities in the discovery of new therapeutics targeting the kynurenine pathway. J Med Chem. 2015;58(22):8762–8782. doi:10.1021/acs.jmedchem.5b00461.
- 29. Miura H, Ozaki N, Sawada M, Isobe K, Ohta T, Nagatsu T. A link between stress and depression: shifts in the balance between the kynurenine and serotonin pathways of tryptophan metabolism and the etiology and pathophysiology of depression. Stress. 2008;11(3):198–209. doi:10.1080/10253890701754068.
- Sforzini L, Nettis MA, Mondelli V, Pariante CM. Inflammation in cancer and depression: a starring role for the kynurenine pathway. Psychopharmacology. 2019;236:2997–3011. doi:10.1007/s00213-019-05200-8.
- Wang Q, Liu D, Song P, Zou MH. Tryptophan-kynurenine pathway is dysregulated in inflammation, and immune activation. Front Biosci (Landmark Ed). 2015;20(7):1116–1143. doi:10.2741/4363.
- Kurgan S, Onder C, Balci N, Akdogan N, Altıngoz SM, Serdar MA, et al. Influence of periodontal inflammation on tryptophankynurenine metabolism: a cross-sectional study. Clin Oral Investig. 2022;26(9):5721–5732. doi:10.1007/s00784-022-04528-4.
- Tonetti MS, Greenwell H, Kornman KS. Staging and grading of periodontitis: Framework and proposal of a new classification and case definition. J Periodontol. 2018;89:S159–S172.
- Onder C, Kurgan S, Altingoz SM, Bagis N, Uyanik M, Serdar MA, et al. Impact of non-surgical periodontal therapy on saliva and serum levels of markers of oxidative stress. Clin Oral Investig. 2017;21:1961–1969. doi:10.1007/s00784-016-1984-z.
- 35. Di Gangi IM, Chiandetti L, Gucciardi A, Moret V, Naturale M, Giordano G. Simultaneous quantitative determination of NG, NG-dimethyl-l-arginine or asymmetric dimethylarginine and related pathway's metabolites in biological fluids by ultrahigh-performance liquid chromatography/electrospray ionization-tandem mass spectrometry. Anal Chim Acta. 2010;677(2):140–148.
- 36. Taylor MW, Feng G. Relationship between interferon- γ , indoleamine 2, 3-dioxygenase, and tryptophan catabolism. FASEB J. 1991;5(11):2516–2522.
- 37. Badawy AAB, Dougherty DM. Assessment of the human kynurenine pathway: comparisons and clinical implications of ethnic and gender differences in plasma tryptophan, kynurenine metabolites, and enzyme expressions at baseline and after acute tryptophan loading and depletion. IJTR. 2016;9:IJTR. S38189. doi:10.4137/Ijtr.S38189.
- 38. Sorgdrager FJ, Naudé PJ, Kema IP, Nollen EA, Deyn PPD. Tryptophan metabolism in inflammaging: from biomarker

to therapeutic target. Front immunol. 2019;10:2565. doi:10.3389/fimmu.2019.02565.

- Choera T, Zelante T, Romani L, Keller NP. A multifaceted role of tryptophan metabolism and indoleamine 2, 3-dioxygenase activity in Aspergillus fumigatus-host interactions. Front immunol. 2018;8:1996. doi:10.3389/fimmu.2017.01996.
- Correale J. Immunosuppressive amino-acid catabolizing enzymes in multiple sclerosis. Front immunol. 2021;11:600428. doi:10.3389/fimmu.2020.600428.
- 41. Zeng Q, Qiu F, Chen Y, Liu C, Liu H, Liang CL, et al. Shikonin prolongs allograft survival via induction of CD4+ FoxP3+ regulatory T cells. Front immunol. 2019;10:652. doi:10.3389/fimmu.2019.00652.
- 42. Hao T, Zhang R, Zhao T, Wu J, Leung WK, Yang J, et al. Porphyromonas gingivalis infection promotes inflammation via inhibition of the AhR signalling pathway in periodontitis. Cell Prolif. 2023;56(2):e13364. doi:10.1111/cpr.13364.
- Palm E, Khalaf H, Bengtsson T. Suppression of inflammatory responses of human gingival fibroblasts by gingipains from Porphyromonas gingivalis. Mol Oral Microbiol. 2015;30(1):74– 85. doi:10.1111/omi.12073.
- 44. Fuchs D, Möller AA, Reibnegger G, Werner ER, Werner-Felmayer G, Dierich MP, et al. Increased endogenous interferongamma and neopterin correlate with increased degradation of tryptophan in human immunodeficiency virus type 1 infection. Immunol Lett. 1991;28(3):207–211. doi:10.1016/0165-2478(91)90005-u.
- Schroecksnadel K, Winkler C, Fuith LC, Fuchs D. Tryptophan degradation in patients with gynecological cancer correlates with immune activation. Cancer Lett. 2005;223(2):323–329. doi:10.1016/j.cca.2005.06.013.
- 46. Barrington G, Khan S, Kent K, Brennan DS, Crocombe LA, Bet-

tiol S. Obesity, dietary sugar and dental caries in Australian adults. Int Dent J. 2019;69(5):383–391. doi:10.1111/idj.12480.

- Khan S, Bettiol S, Kent K, Barnett T, Peres M, Crocombe LA. Obesity and periodontitis in Australian adults: a populationbased cross-sectional study. Int Dent J. 2020;70(1):53–61. doi:10.1111/idj.12514.
- Atabay V, Lutfioğlu M, Avci B, Sakallioglu E, Aydoğdu A. Obesity and oxidative stress in patients with different periodontal status: a case-control study. J Periodontal Res. 2017;52(1):51–60. doi:10.1111/jre.12368.
- 49. Nascimento GG, Leite FR, Correa MB, Peres MA, Demarco FF. Does periodontal treatment have an effect on clinical and immunological parameters of periodontal disease in obese subjects? A systematic review and meta-analysis. Clin Oral Investig. 2016;20:639–647.
- Ylöstalo P, Suominen-Taipale L, Reunanen A, Knuuttila M. Association between body weight and periodontal infection. J Clin Periodontol. 2008;35(4):297–304.
- Kang J, Smith S, Pavitt S, Wu J. Association between central obesity and tooth loss in the non-obese people: results from the continuous National Health and Nutrition Examination Survey (NHANES) 1999–2012. J Clin Periodontol. 2019;46(4):430–437.
- Palle AR, Reddy CSK, Shankar BS, Gelli V, Sudhakar J, Reddy KKM. Association between obesity and chronic periodontitis: a cross-sectional study. J Contemp Dent Pract. 2013;14(2):168.
- 53. Mangge H, Summers KL, Meinitzer A, Zelzer S, Almer G, Prassl R, et al. Obesity-related dysregulation of the Tryptophan–Kynurenine metabolism: Role of age and parameters of the metabolic syndrome. Obesity. 2014;22(1):195–201. doi:10.1002/oby.20491.